



Informing Drug Design: Human Acetylcholinesterase Response to Organophosphate Poisoning

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Abstract

Acetylcholinesterase (AChE) is a target enzyme of organophosphate (OP). Current treatments for OP poisoning, i.e. oximes, have limited success, especially without pre-treatment. This study uses molecular dynamic analysis to shine light on structure and dynamical fluctuations of free AChE and OP-inhibited AChE. Knowledge gained by the study of OP inhibition of acetylcholinesterase should guide future drug designs of more effective antitoxins.

Background

- AChE is a serine protease that breaks down the neurotransmitter acetylcholine to terminate neurotransmission.
- AChE is present in all nerve synapses, neuromuscular junctions, and RBCs.
- OPs are commonly found oil additives, pesticides, and chemical weapons, which can target and inhibit AChE.

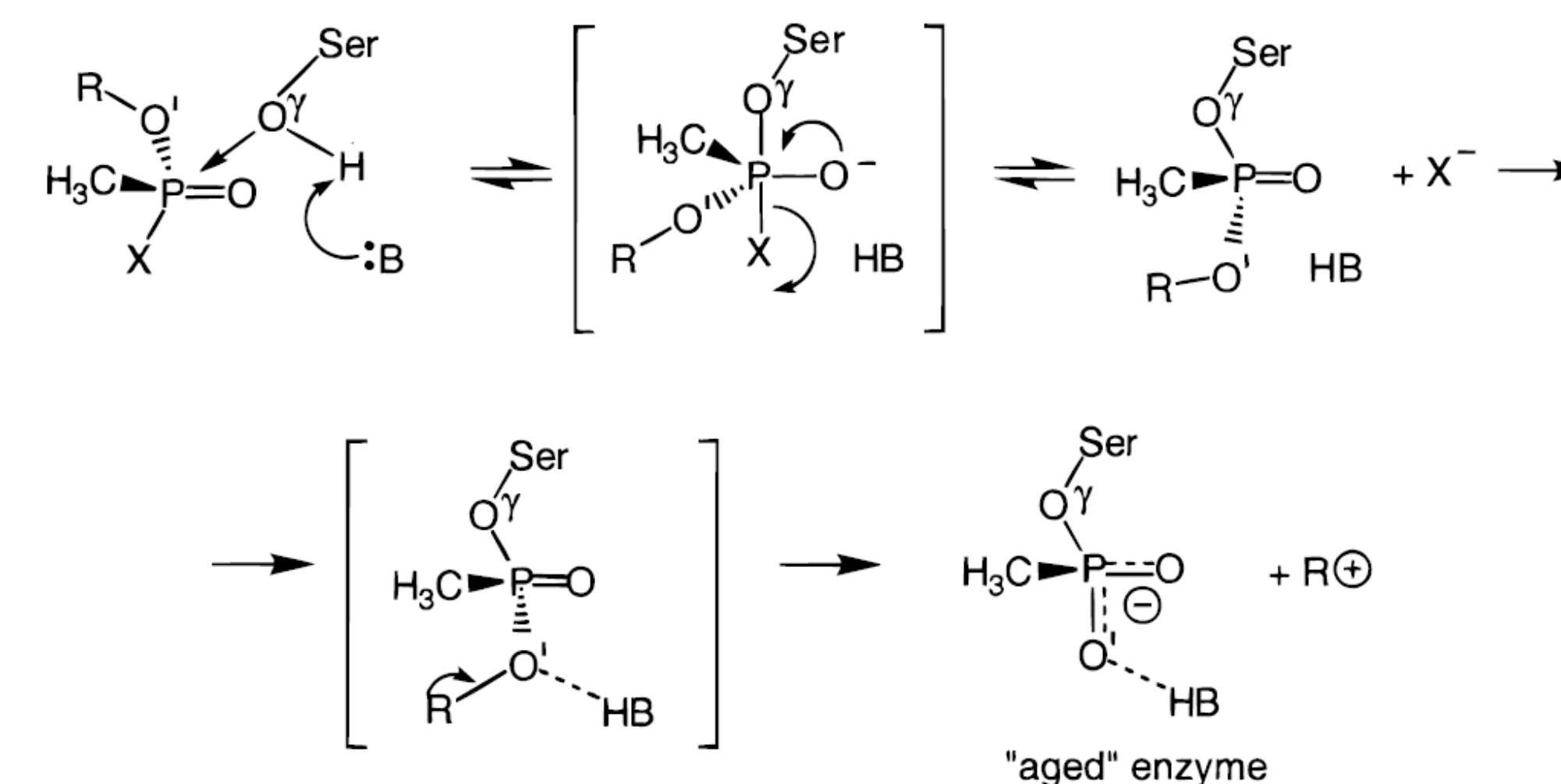


Figure 1. Proposed mechanism for irreversible "aging" of AChE by an OP.

- Acute cholinergic crisis, is the major manifestation of OP poisoning. Inhibition of synaptic AChE causes an accumulation of acetylcholine in the nerve synapse leading to continuous neurotransmission and possible death within minutes.
- Molecular Dynamic (MD) analysis was performed to elucidate characteristics of enzyme/adduct to aid design of more effective countermeasures of OP poisoning.

Methods

1

NAMD, Scalable Molecular Dynamics: Parallel molecular dynamics code for high-performance simulation

2

Essential Dynamics Analysis: Covariance matrix of positional fluctuations of the Ca atoms analyzed reveal principle directions of large concerted motions used to assess dynamical similarity

3

VMD, Visual Molecular Dynamics: displaying, animating and analyzing biomolecular systems (Solvent Accessible Surface Area, Distance Plots, Modes of Motion Comparison)

4

MOLE: Rapid and fully automated location and characterization of channels, tunnels, and pores in molecular structures

Results and Conclusions

1

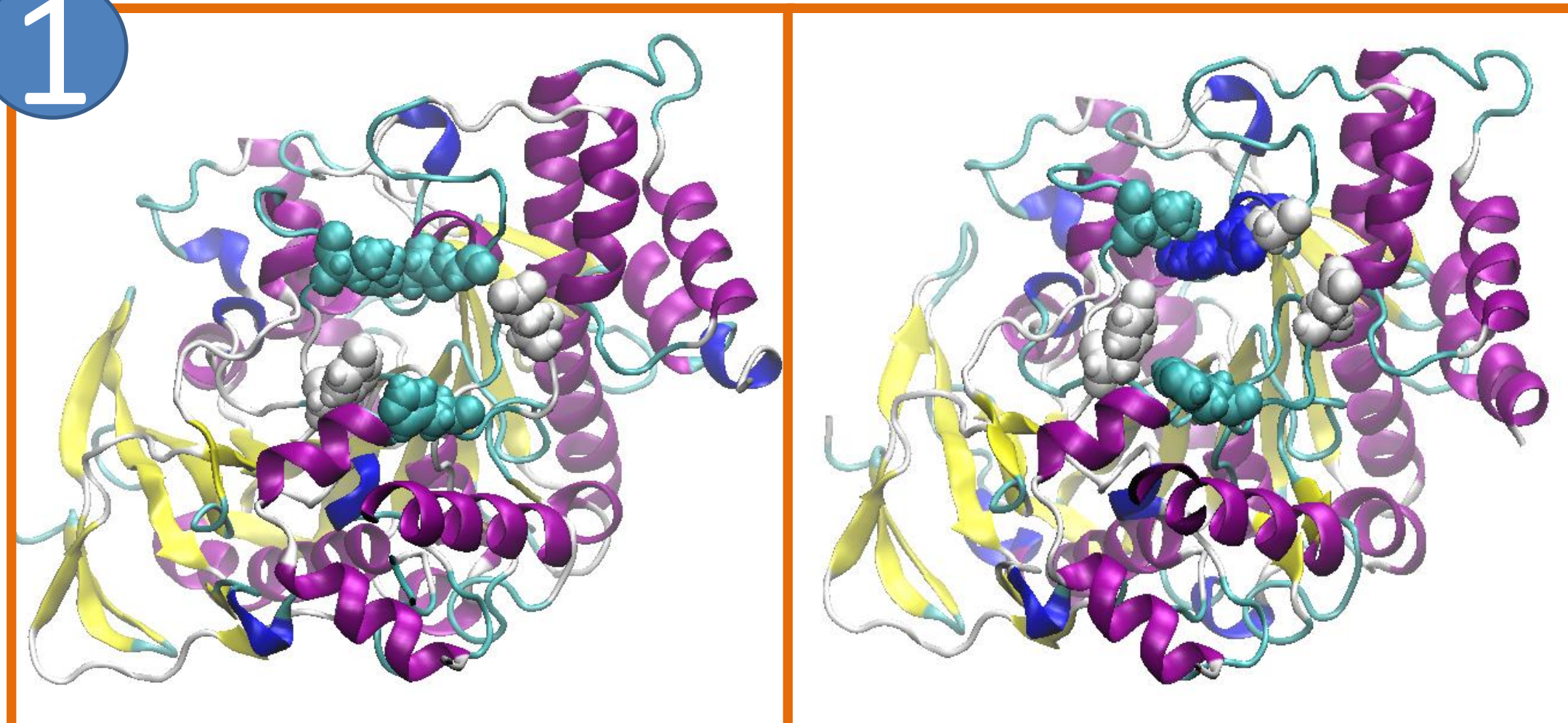


Figure 2. Beginning structure of apoprotein simulation colored by secondary structure.

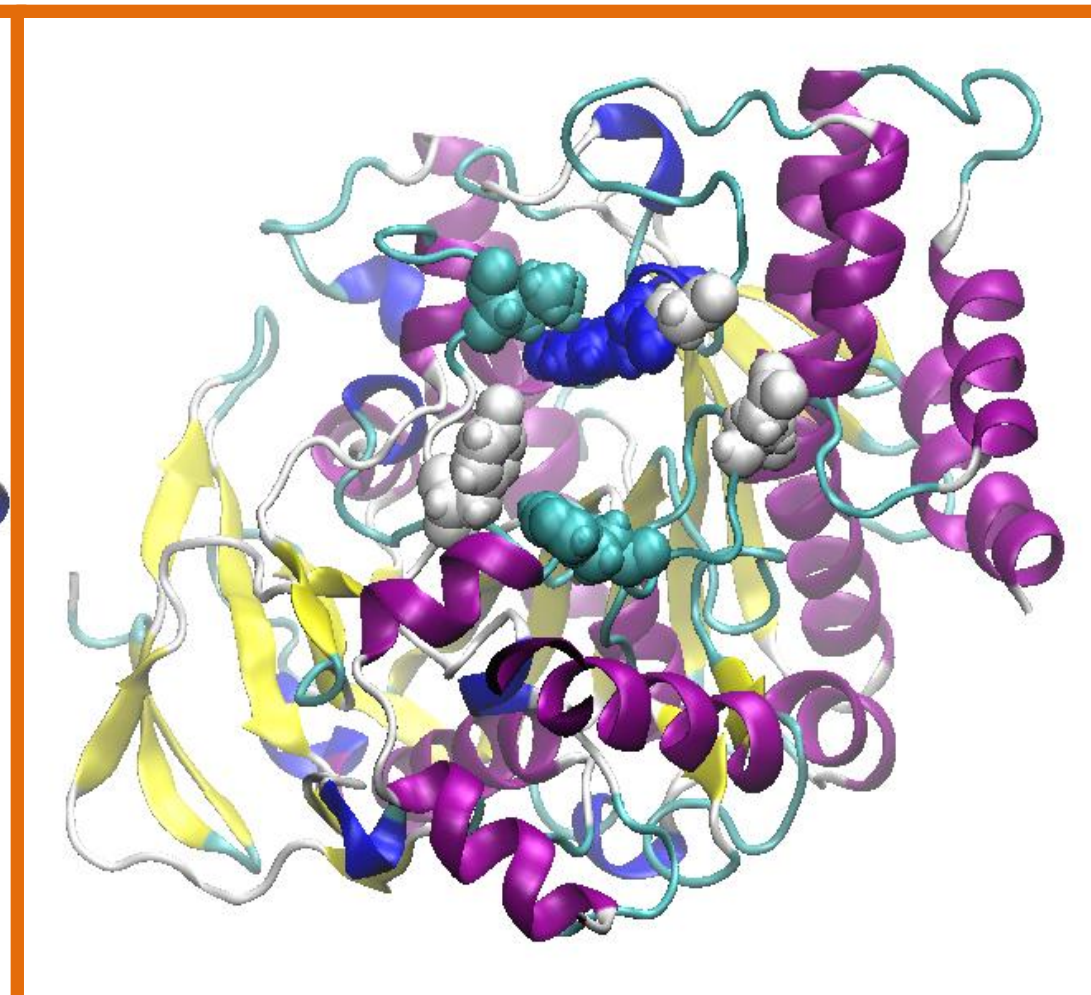


Figure 3. Beginning Structure of soman-adducted simulation colored by secondary structure.

2

	APO-AChE	Soman-AChE
APO-AChE	0.628	0.362
Soman-AChE		0.527

Table 1. Root-Mean Square Inner Product (RMSIP) calculations comparing non-adducted simulation to OP-adducted simulations.

- Minimal conformational differences between initial structures of simulations.
- Simulations adequately sampled essential subspace. Apoprotein simulation compared with soman-adducted simulation show low similarity of overall motions.

3

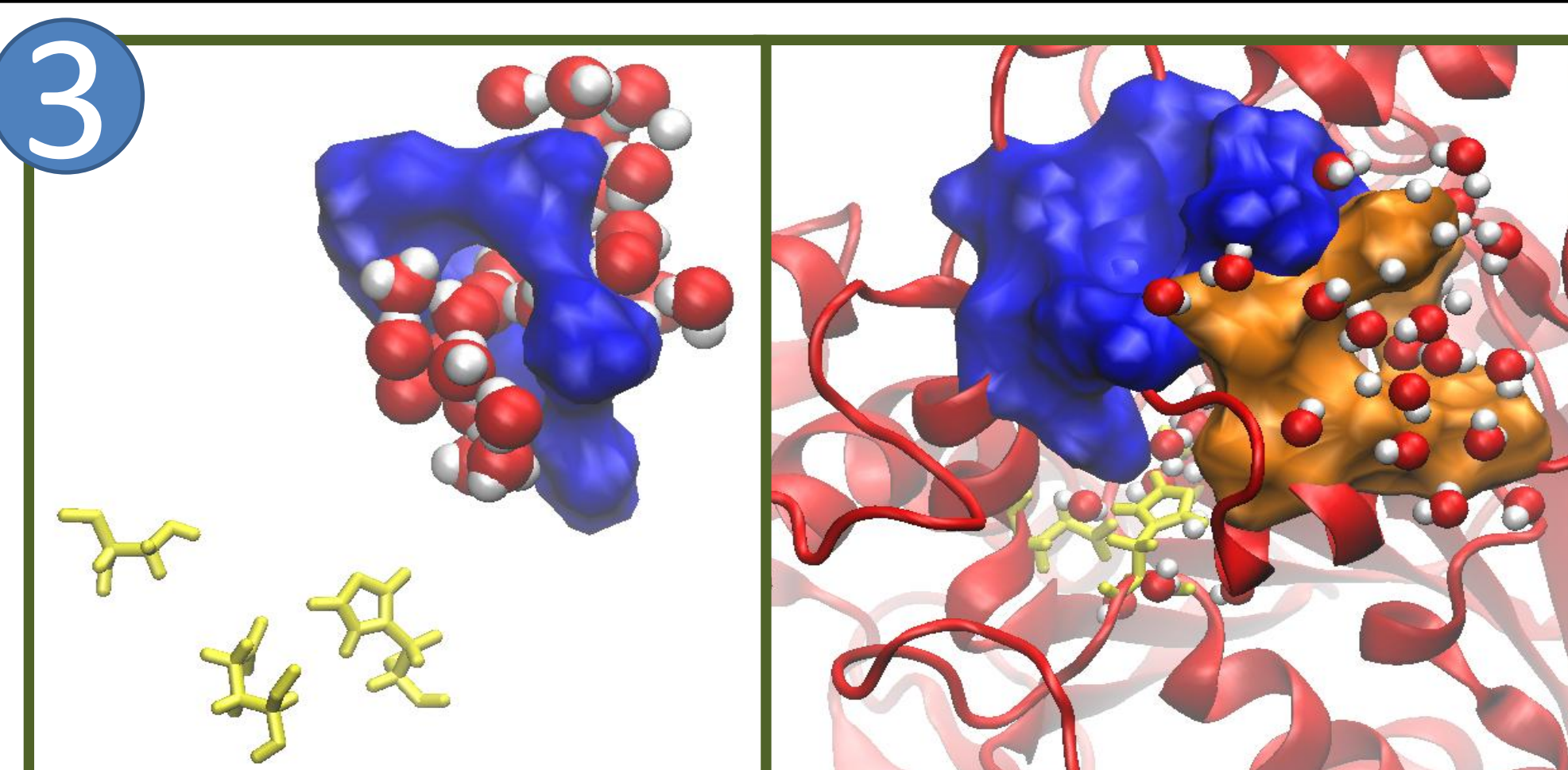


Figure 4. Entrance to Active Site of Apoprotein. In blue, main gorge opening. In yellow, catalytic triad of active site.

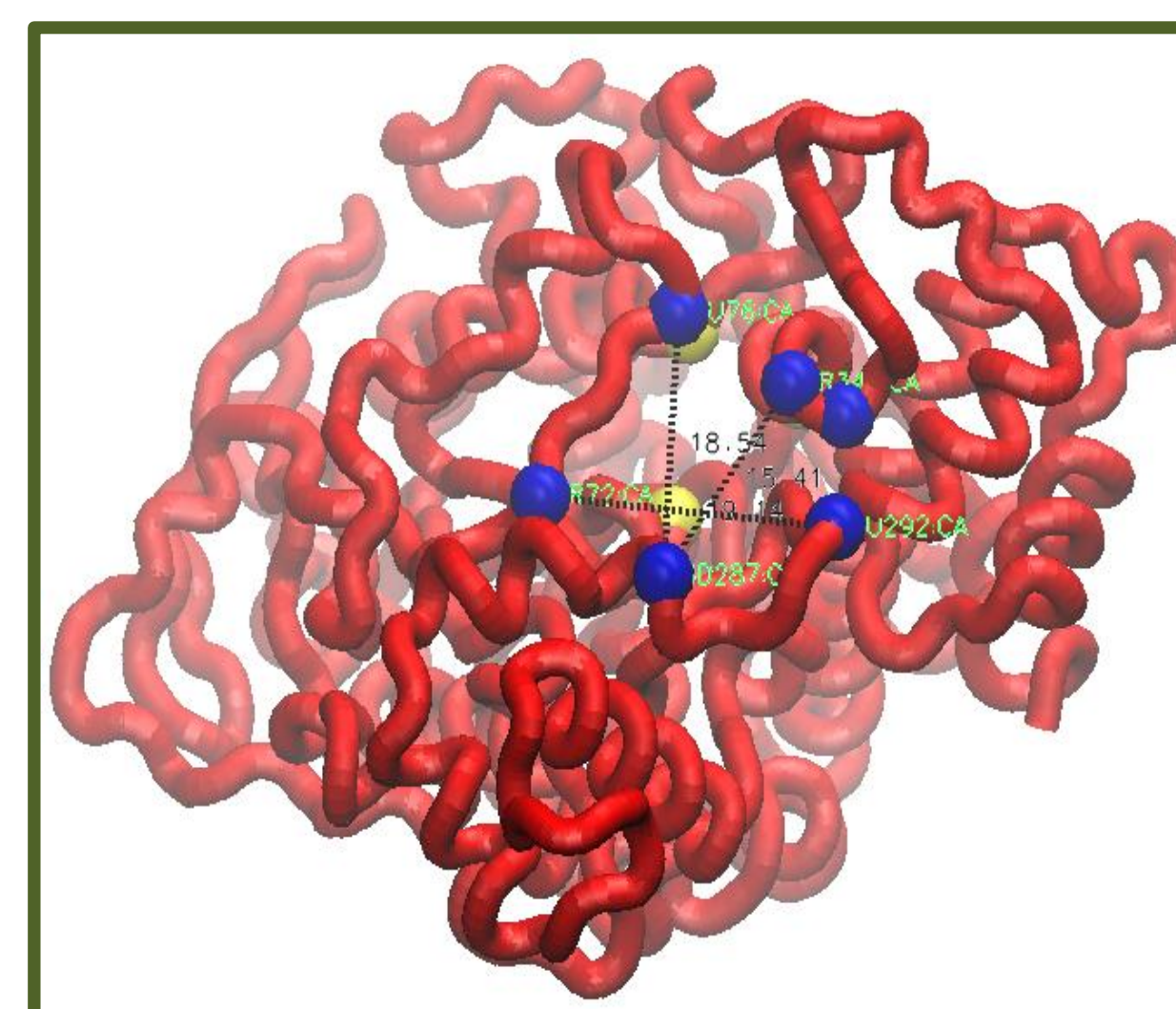


Figure 5. Side Entrance to Active Site of Apoprotein. In orange, side door opening. In blue, main gorge closed. In yellow, catalytic triad of active site.

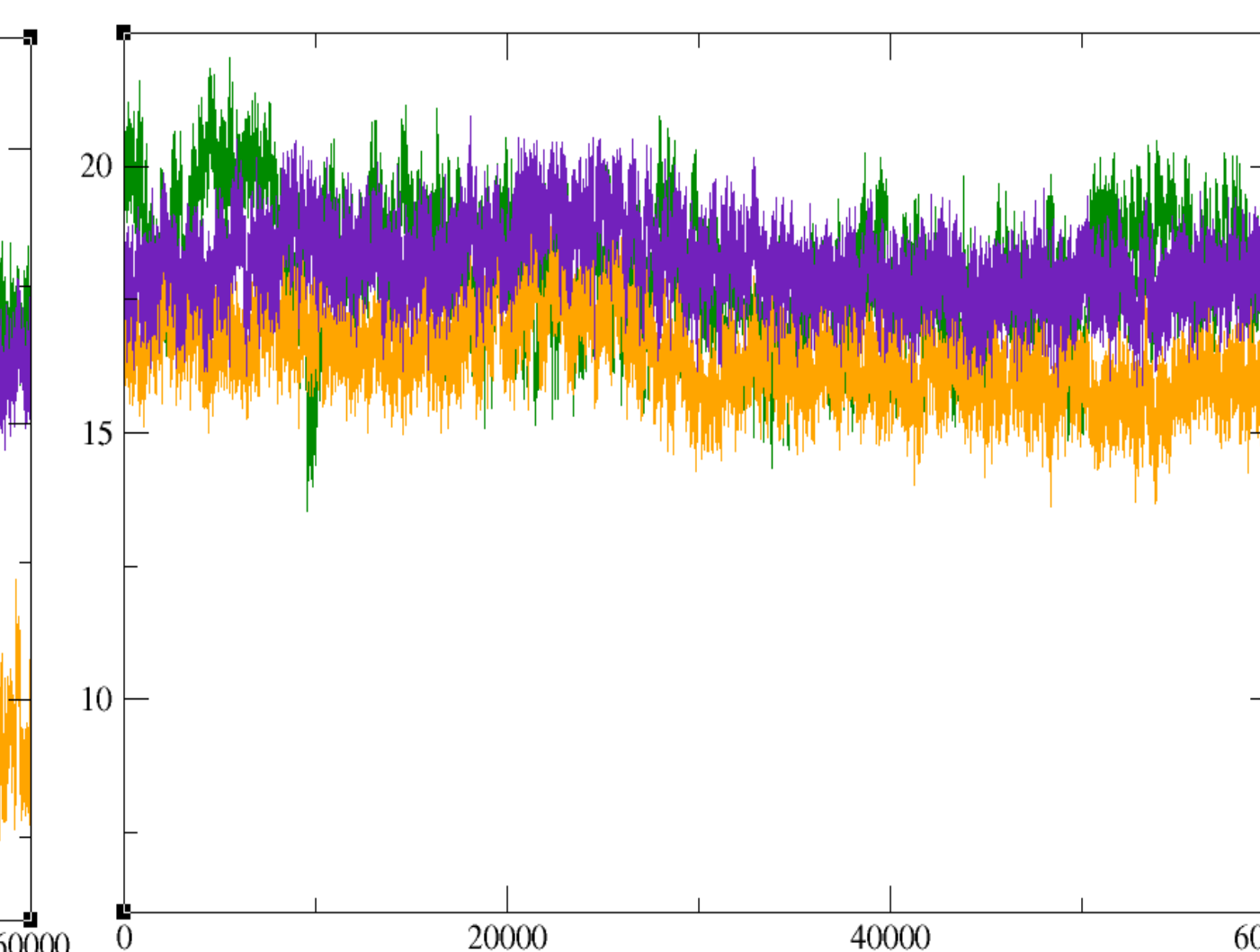
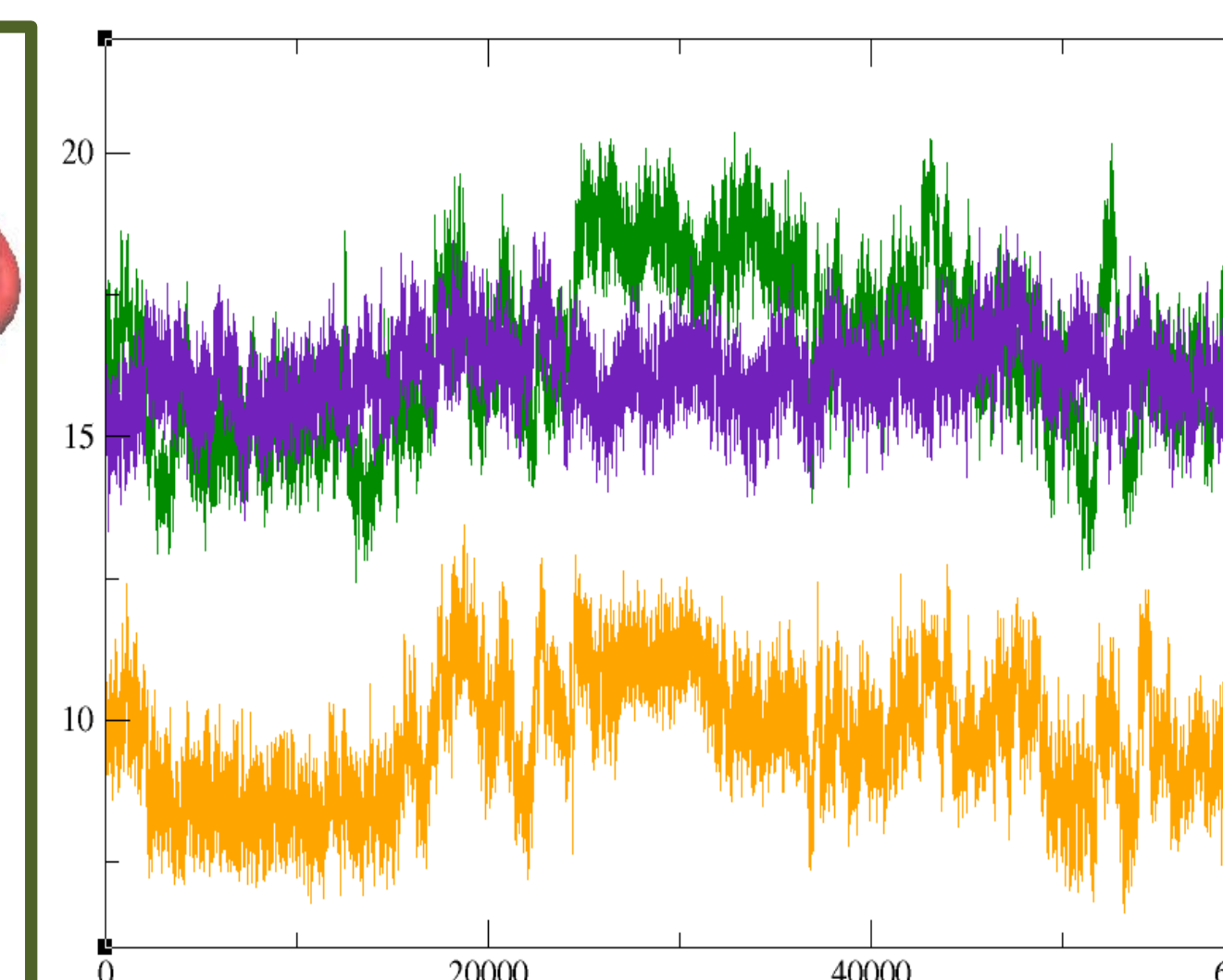


Figure 6. Main door opening in soman-adducted simulation. In blue, isolated residues of main door. In yellow, catalytic triad of active site.

Figure 7. Distance plot across gorge of apoprotein simulation reveals more drastic gorge fluctuations, supported by visual observation of principle modes of movement.

Figure 8. Distance plot across gorge of adducted simulation reveals rigid gorge fluctuations, supported by visual observation of principle modes of movement.

4

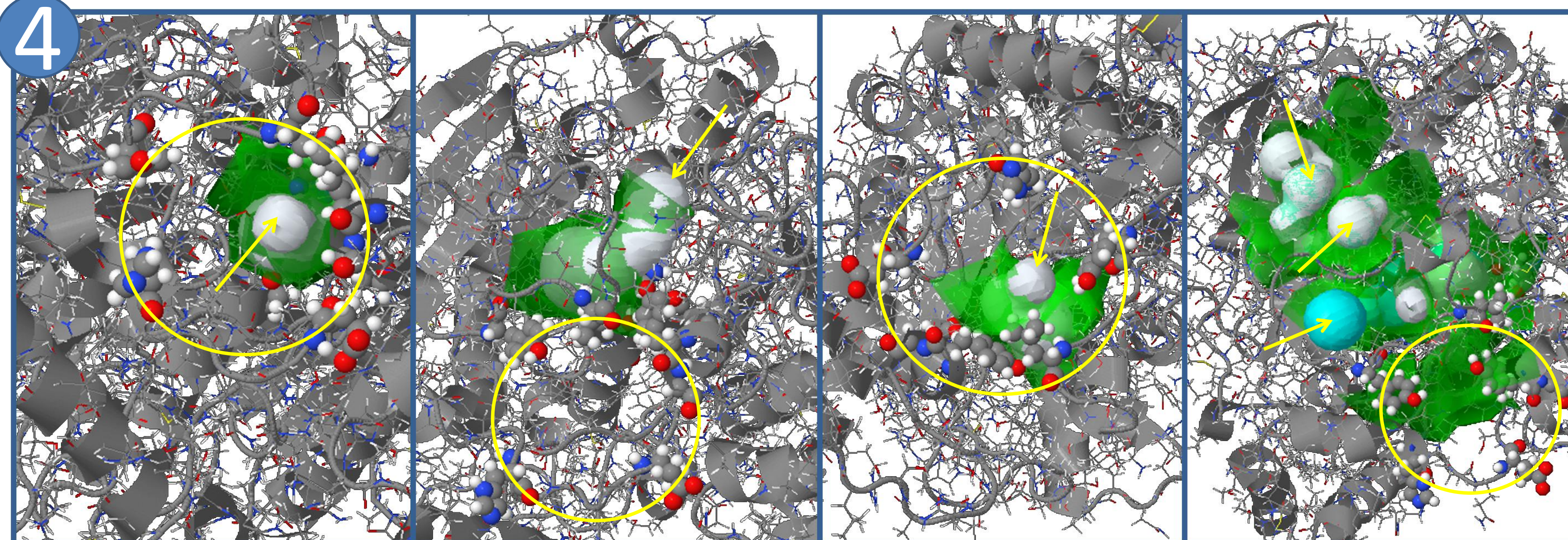


Figure 9. Tunnel predicted by MOLE corresponds to main gorge in apoprotein. Cavity (green) volume: 483 Å³

Figure 10. Tunnel predicted by MOLE reveals opened side door in apoprotein. Cavity (green) volume: 704 Å³

Figure 11. Tunnel predicted by MOLE corresponds to main gorge in adducted protein. Cavity (green) volume: 483 Å³

Figure 12. Tunnels predicted by MOLE reveal opened side and back doors while main gorge opening is closed in adducted protein. Cavity (green) volume: 1351 Å³

- MOLE analysis reveals side doors open while main gorge is closed and also suggests gorge volume is not necessarily correlated with main door configuration.

References

- Ariel, N., Orndorff, A., Barak, D., Bino, T., Velan, B., & Shafferman, A. (1998). The 'aromatic patch' of three proximal residues in the human acetylcholinesterase active centre allows for versatile interaction modes with inhibitors. *Biochemical Journal*, 335, 95-102.
- Bossa, C., Amadei, A., Daidone, I., Anselmi, M., Vallone, R., Brunori, M., & Di Nola, A. (2005). Molecular dynamics simulation of sperm whale myoglobin: Effects of mutations and trapped CO on the structure and dynamics of cavities. *Biophysical Journal*, 89(1), 465-474.
- Clement, J., Ehrhard, N. (1994). In vitro oxime-induced reactivation of various molecular forms of soman-inhibited acetylcholinesterase in striated muscle from rat, monkey and human. *Archives of Toxicology*, 68(10), 686-695.
- Colletier, J. P., Royant, A., Specht, A., Sanson, B., Nachon, F., Masson, P., ... Weik, M. (2007). Use of a 'caged' analogue to study the traffic of choline within acetylcholinesterase by kinetic crystallography. *Acta Crystallographica Section D: Biological Crystallography*, 63, 1115-1128.
- de Koning, M. G., van Groen, M., & Scott, D. (2011). Peripheral site ligand conjugation to a non-quaternary oxime enhances reactivation of nerve agent-inhibited human acetylcholinesterase. *Toxicology Letters*, 206(1), 54-59.
- Eckert, S., Eyer, P., Muckler, H., & Worek, F. (2006). Kinetic analysis of the protection afforded by reversible inhibitors against irreversible inhibition of acetylcholinesterase by highly toxic organophosphorus compounds. *Biochemical Pharmacology*, 72(3), 344-357.
- Eyreedy, J. J., Kovach, I. M., & Bencusa, A. (2001). Molecular dynamics study of active-site interactions with tetrahedral intermediates in acetylcholinesterase and its mutants. *Biochemical Journal*, 353, 645-653.
- Fang, J., Pan, Y. M., Murty, J. L., & Zhao, C. G. (2011). Active Site Gating and Substrate Specificity of Butyrylcholinesterase and Acetylcholinesterase: Insights from Molecular Dynamics Simulations. *Journal of Physical Chemistry B*, 115(27), 8797-8805.
- Humphrey, W., Dalke, A., and Schulten, K. (1996) VMD - Visual Molecular Dynamics. *J. Mol. Graphics*, 14(1), 33-38.
- Hsieh, B. H., Deng, J. F., Gu, J., & Tsai, W. J. (2003). Acetylcholinesterase inhibition and the extrapyramidal syndrome: A review of the neurotoxicity of organophosphate. [Review]. *Neurotoxicology*, 24(4), 423-427.
- Kua, J., Zhang, Y. K., Ismaili, A. C., Butler, J. R., & McCammon, J. A. (2003). Studying the roles of W86, E202, and Y337 in binding of acetylcholine to acetylcholinesterase using a combined molecular dynamics and multiple docking approach. *Protein Science*, 12(12), 2675-2684.
- Petek, M., Kostova, P., Koca, J., Olyepka, M. (2007) MOLE: A Voronoi Diagram-Based Explorer of Molecular Channels, Pores, and Tunnels. *Structure*, 15, 1357-1363.
- Ramanathan, A., & Agarwal, P. K. (2011). Evolutionarily Conserved Linkage between Enzyme Fold, Flexibility, and Catalysis. *Plos Biology*, 9(11).
- Sanson, B., Colletier, J. P., Xu, Y. C., Lang, P. T., Jiang, H. L., ... Weik, M. (2011). Backdoor opening mechanism in acetylcholinesterase based on X-ray crystallography and molecular dynamics simulations. *Protein Science*, 20(7), 1114-1118.
- Worek, F., Szinicz, L., & Thiermann, H. (2005). Estimation of oxime efficacy in nerve agent poisoning: A kinetic approach. *Chemico-Biological Interactions*, 157, 349-352.
- Worek, F., Wille, T., Koller, M., & Thiermann, H. (2012). Reactivation kinetics of a series of related bispyridinium oximes with organophosphate-inhibited human acetylcholinesterase: Structure-activity relationships. *Biochemical Pharmacology*, 83(2), 1700-1706.
- Xu, Y. C., Colletier, J. P., Jiang, H. L., Siman, J., Sussman, J. L., & Weik, M. (2008). Induced-fit or preexisting equilibrium dynamics? Lessons from protein crystallography and MD simulations on acetylcholinesterase and implications for structure-based drug design. *Protein Science*, 17(4), 601-605.

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