Nutrient Uptake and Organic Acid Anion Metabolism in Lupins and Peas Supplied with Nitrate

S. P. LOSS, A. D. ROBSON and G. S. P. RITCHIE

Unlike many plants reported in the literature, lupins do not excrete OH⁻ in amounts equivalent to the net excess of inorganic anion uptake over inorganic cation uptake. To investigate the mechanisms involved in the maintenance of charge balance, nutrient uptake and organic anion accumulation of lupins and peas supplied with a range of NO₃⁻ concentrations, were compared.

Lupins absorbed less NO₃⁻ than peas on a dry weight basis, which largely accounted for the smaller excess of anion uptake over cation uptake in lupins than in peas at the same NO₃ supply. When anion uptake exceeded cation uptake, peas excreted an equivalent charge of OH⁻ whereas lupins excreted much smaller amounts of OH⁻ than the excess of anion over cation uptake. It was calculated that lupins excreted significant amounts of organic anions when anion uptake exceeded cation uptake, whereas organic anion excretion from peas was negligible, regardless of their NO₃ supply and cation–anion balance.

In this study, organic anion excretion was measured from lupin roots grown in near-sterile conditions while supplied with NO₃⁻ at 0, 500 and 2000 μM. Although complete sterility was not achieved, there was close agreement between the organic anion excreted and the excess anion over cation uptake.

INTRODUCTION

Unlike most plants (Van Beusichem, 1981; Jarvis and Robson, 1983; Van Beusichem, Kirkby and Baas, 1988), nodulated lupins (Lupinus angustifolius L.) grown in solution culture did not excrete OH⁻ when supplied with NO₃ (Loss, Robson and Ritchie, 1993b). When NO₃ was supplied to the lupins at 750 μM, inorganic cation–anion uptake was balanced but H⁺ excretion continued to occur, and when NO₃ was supplied at 5000 μM, inorganic anion uptake exceeded inorganic cation uptake and the pH of the nutrient solution was unaffected. Similar results were obtained by Atwell (1992), who found that the pH of the nutrient solution did not increase when lupins were supplied with NO₃⁻.

To explain these results, we proposed that lupins maintain their internal charge balance by excreting organic acid anions rather than OH⁻ in exchange for the uptake of inorganic anions (Loss et al., 1993b). Malate and citrate are the organic anions present in the largest quantities in lupins and their pKa values are low enough not to affect the external pH of the growing medium at pH 6-7. The excretion of H⁺ coupled to cation uptake and the reduction of NO₃⁻ and SO₄²⁻ within plants produces OH⁻ and any rise in cellular pH is prevented by the synthesis of organic acids (Davies, 1973). Greater organic anion accumulation has been measured in castor oil plants (Ricinus communis L.) supplied with NO₃⁻ than in those supplied with NH₄⁺ (Van Beusichem et al., 1988). Unlike many plants, including peas (Pisum sativum L.), the reduction of NO₃⁻ is mainly confined to the roots of lupins when supplied with 5 mM NO₃ or less, and it is only at high NO₃ supplies (10 mM) that significant quantities are reduced in the shoots (Andrews et al., 1984). This could indicate a difference in the mechanisms of charge balance.

The aim of this study was to investigate nutrient uptake and organic anion metabolism in lupins and peas supplied with a range of NO₃ concentrations.

MATERIALS AND METHODS

Experiment 1

Experimental design. The aim of this experiment was to compare nutrient uptake and organic anion accumulation in unnodulated peas and lupins supplied with NO₃⁻, with the hypothesis that peas absorb more NO₃⁻ and accumulate more organic anions than lupins. The effects of four concentrations of NO₃⁻ (250, 750, 2500, 5000 μM) on nutrient uptake, cation–anion balance, H⁺/OH⁻ excretion and organic anion accumulation in the roots and shoots of unnodulated lupin and pea seedlings were examined. The experiment included three replicates and was conducted in root cooling tanks at 20 °C, in an air-conditioned glasshouse during Sep. and Oct. 1990, when internal air temperatures varied between 15 and 25 °C.

Seedling preparation. Seeds of L. angustifolius cv. Yandee and P. sativum L. cv. Dundale were sterilised with a 1%
solution of NaOCl to prevent any contamination with rhizobia. The seeds were then germinated on a stainless steel screen suspended on the surface of an aerated solution of 10^-4 M CaSO4 and 10^-6 M H3BO3. After 7 d, seedlings were transferred to an aerated, nutrient solution at a density of eight seedlings per 50 l pot. The nutrient solution contained the following concentrations of nutrients (μM): CaSO4, 625; K2SO4, 600; MgSO4, 200; NaH2PO4, 30; H3BO3, 5; FeNaEDTA, 3; MnSO4, 10; ZnSO4, 0.75; CuSO4, 0.2; CoSO4, 0.2; Na2MoO4, 0.03. Apart from N, the solution provided an adequate but not excessive nutrient supply to the young seedlings. The pH of the solution was maintained between 4.5 and 6.5 with daily additions of 0.1 M KOH and the solutions were changed every second day. After 3 weeks, the small number of plants that had formed nodules on their roots were discarded and the seedlings were showing the first symptoms of N deficiency.

Treatments. Thirty six lupin and pea seedlings were transferred to the treatments pots at a density of three plants per 50 l pot. In the 5000 μM NO3 treatment, CaSO4 was withheld and NO3 was supplied as Ca(NO3)2. In the 250, 750 and 2500 μM NO3 treatments, ratios of Ca(NO3)2 and CaSO4 were supplied so that Ca2+ concentration was 2500 μM in all treatments. Control pots without plants were included for each treatment. pH was monitored in each pot four times daily, and corrected to 5.7 with 0.01 M NaOH or H3BO3. The pH of the solution was maintained constant pH during the treatment period. The nutrient solution was passed through a No. 1 Whatman filter paper and allowed to dry at room temperature for 16–32 h. The sample was then dissolved in 20 ml of double de-ionised water, to which about 20 ml of petroleum ether (b.p. < 40°C) was added. The sample was hand shaken and allowed to stand several times and was then placed in a freezer. When the aqueous phase had frozen the liquid ether phase containing the organic soluble components of the sample was discarded and the aqueous phase was allowed to thaw. The addition to ether, shaking and freezing the sample was repeated another two times. Using a Waters® ion chromatography system, 100 μl of each sample was injected into an Aminex® ion exclusion column (HPX-87H). The eluent was 2.5 mM H3PO4 at a flow rate of 0.8 ml min^-1 and the organic anions were detected by a uv/vis spectrophotometer at 210 nm.

Inorganic cation-anion balance, H+ excretion and organic anion accumulation were expressed per g dry weight per d. The amount of organic anions excreted was estimated from the following equations. The charge of the anions and cations absorbed must equal the charge of the anions and cations excreted:

\[ A_{\text{absorbed}} - C_{\text{absorbed}} = OA_{\text{excreted}} + OH_{\text{excreted}} \]

where A and C are inorganic anions and cations (other than H+ or OH-) respectively, and OA is organic acid anions. Hence, organic anion excretion was calculated using:

\[ OA_{\text{excreted}} = (A_{\text{absorbed}} - C_{\text{absorbed}}) - OH_{\text{excreted}} \]

Experiment 2

The aim of this experiment was to measure the effects of three concentrations of NO3 (0, 500 and 2000 μM) on nutrient uptake, cation-anion balance, H+/OH- excretion and organic anion accumulation and excretion of unnodulated lupin plants. The experiment included six replicates and was conducted in a controlled growth room at 20°C and a 12 h photoperiod, during Nov. and Dec. 1991.

Seedling preparation. For expt 2, an attempt was made to prepare seedling roots in sterile conditions, because a preliminary experiment demonstrated that at the expected rate of excretion, the organic anions were decomposed within 48 h. All equipment and nutrient solutions were autoclaved at 121°C for 20 min and were handled in a laminar flow cabinet once sterile. Seeds were surface sterilised with 12% NaOCl for 4 h, rinsed ten times with excess sterile de-ionised water and allowed to imbibe overnight in a sterile solution of 10^-4 M CaSO4 and 10^-6 M H3BO3. The seeds were then immersed in a 3% solution of H2O2 for 10 s, rinsed and transferred to sterile 500 ml screw top polycarbonate vials containing sterile nutrient solution. The solution was aerated through a sterile Pasteur pipette packed with cotton wool and the seeds were supported on a stainless steel mesh at a density of 20 per vial. The composition of the nutrient solution was as in expt 1, but also included 30 mg l^-1 of the antibiotic ceftoxime (Kerven et al., 1991), which did not decrease lupin growth and 250 μM NaN3. The solution was changed every second day. Preliminary tests showed that contamination was not prevented by 50 mg ml^-1 ampicillin and 50 mg ml^-1 strep-
tomycin, at which concentration the growth of plants was decreased.

After 1 week, the seedling shoots were exposed to non-sterile external air through a hole in the lid of the vial, while the roots remained in the sterile nutrient solution. The seedlings were supported by sterile cotton wool around the hole in the lid with two seedlings per vial. After 3 weeks cotyledons were excised to ensure the plants were relying solely on the nutrient solution supplied. Throughout the pretreatment period, sterility was tested weekly. A small sample of nutrient solution was plated on yeast mannitol agar and examined after incubation for 48 h at 20°C.

**Treatments.** The treatments were imposed when the plants were 4 weeks old. In the 500 and 2000 μM NO₃⁻ treatments, NO₃⁻ was supplied as Ca(NO₃)₂ and ratios of Ca(NO₃)₂ and CaSO₄ were supplied so that the Ca²⁺ concentration was 1000 μM in all treatments. The supply of K₂SO₄, MgSO₄, NaH₂PO₄ and the micronutrients was doubled from those of the pretreatment, because the volume of nutrient solution per plant was much less than in expt 1. Citrate (100 μg l⁻¹) was also added to six pots with plants from which NO₃⁻ was withheld, so that any loss from the system due to microbial decomposition could be measured. The treatment solutions were replaced after 4 d and again after another 4 d before the plants were harvested. After each treatment period, samples were collected and the nutrient solutions were titrated back to their initial pH.

**Analyses.** Nutrient uptake, cation–anion balance, H⁺ excretion and organic anion accumulation were calculated as described as in expt 1 and expressed per g dry weight per d. The concentrations of organic anions in the nutrient solution were determined using ion chromatography as described previously, apart from the eluent, which was changed to 4 mM H₂SO₄ at a flow rate of 0.6 ml min⁻¹.

**RESULTS**

**Experiment 1**

The growth of the seedlings during the pre-treatment phase was as expected, with the peas showing the first symptoms of N deficiency. These symptoms persisted throughout the treatment period for the 250 μM NO₃⁻ treatment. For the other treatments there was no difference (P < 0.01) in the effect of the treatments on shoot or root growth for either the lupins or peas (Table 1).

**Nutrient uptake and cation–anion balance.** Unlike the other nutrients NO₃⁻ was totally depleted in the 250 μM NO₃⁻ treatment of both the peas and lupins, and there was a net

<table>
<thead>
<tr>
<th>NO₃⁻ treatment (μM)</th>
<th>250</th>
<th>750</th>
<th>2500</th>
<th>5000</th>
<th>l.s.d.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lupin Roots</td>
<td>0.91</td>
<td>0.81</td>
<td>0.95</td>
<td>0.80</td>
<td>0.16</td>
</tr>
<tr>
<td>Shoots</td>
<td>0.69</td>
<td>0.52</td>
<td>0.56</td>
<td>0.55</td>
<td>0.12</td>
</tr>
<tr>
<td>Pea Roots</td>
<td>0.42</td>
<td>0.51</td>
<td>0.56</td>
<td>0.55</td>
<td>0.12</td>
</tr>
<tr>
<td>Shoots</td>
<td>0.72</td>
<td>0.91</td>
<td>0.99</td>
<td>0.86</td>
<td>0.15</td>
</tr>
</tbody>
</table>

**TABLE 1. The mean root and shoot dry weights (g) of the various treatments and l.s.d. (P < 0.01) in expt 1**

<table>
<thead>
<tr>
<th>N Treatment (μM)</th>
<th>K⁺</th>
<th>Na⁺</th>
<th>Ca²⁺</th>
<th>Mg²⁺</th>
<th>H₂PO₄⁻</th>
<th>SO₄²⁻</th>
<th>Cl⁻</th>
<th>NO₃⁻</th>
<th>C⁻</th>
<th>A⁻</th>
<th>Calculated OA accumulation* over the 6 d treatment period µeq g⁻¹ d⁻¹</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lupin 750</td>
<td>67</td>
<td>53</td>
<td>249</td>
<td>32</td>
<td>9</td>
<td>529±41</td>
<td>549±12</td>
<td>20</td>
<td>180±8</td>
<td>180±8</td>
<td>180±8</td>
</tr>
<tr>
<td>Shoots 750</td>
<td>27</td>
<td>22</td>
<td>249</td>
<td>32</td>
<td>9</td>
<td>529±41</td>
<td>549±12</td>
<td>20</td>
<td>180±8</td>
<td>180±8</td>
<td>180±8</td>
</tr>
<tr>
<td>Pea 750</td>
<td>22</td>
<td>18</td>
<td>249</td>
<td>32</td>
<td>9</td>
<td>529±41</td>
<td>549±12</td>
<td>20</td>
<td>180±8</td>
<td>180±8</td>
<td>180±8</td>
</tr>
<tr>
<td>Pea 2500</td>
<td>89</td>
<td>11</td>
<td>196</td>
<td>27</td>
<td>8</td>
<td>939±26</td>
<td>939±26</td>
<td>20</td>
<td>180±8</td>
<td>180±8</td>
<td>180±8</td>
</tr>
<tr>
<td>Pea 5000</td>
<td>81</td>
<td>18</td>
<td>249</td>
<td>32</td>
<td>9</td>
<td>939±26</td>
<td>939±26</td>
<td>20</td>
<td>180±8</td>
<td>180±8</td>
<td>180±8</td>
</tr>
</tbody>
</table>

* Standard errors.
Fig. 1. The relationship between cation-anion balance and $H^+$ excretion for (□) lupins and (○) peas in expt 1. Individual replicates are illustrated.

Fig. 2. The proportion of organic anions ([■] malate, [□] citrate, [□] others] found in the roots and shoots of lupin and peas supplied with 250 or 5000 $\mu$M NO$_3$ in expt 1.

Table 3. The root and shoot dry weights (g) of the treatments in expt 2. No differences were significant ($P < 0.01$).

<table>
<thead>
<tr>
<th>NO$_3$ Treatments ($\mu$M)</th>
<th>0</th>
<th>500</th>
<th>2000</th>
<th>1.s.d.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Roots</td>
<td>0.47</td>
<td>0.42</td>
<td>0.43</td>
<td>0.09</td>
</tr>
<tr>
<td>Shoots</td>
<td>0.54</td>
<td>0.50</td>
<td>0.52</td>
<td>0.14</td>
</tr>
</tbody>
</table>

cation excess (Table 2). Apart from Na$^+$, Mg$^{2+}$ and Cl$^-$, the peas absorbed more nutrients than the lupins, particularly SO$_4^{2-}$ and NO$_3^-$. These anions contributed to the trend of greater excess anion uptake with increasing NO$_3^-$ supply for both species, particularly the peas.

$H^+$/OH$^-$ excretion. The peas increased the pH of the nutrient solution when supplied with greater than 250 $\mu$M NO$_3^-$, and there was a close linear relationship between the cation–anion balance and the amount of $H^+$/OH$^-$ required to maintain a constant pH in the nutrient solution ($r^2 = 0.98$, Fig. 1). When the lupins absorbed an excess of anions, they excreted less than 190 $\mu$eq OH$^-$ g$^{-1}$ d$^{-1}$ into the nutrient medium, even when anion uptake exceeded cation uptake by 920 $\mu$eq g$^{-1}$ d$^{-1}$.

Organic anion accumulation. The total amount of organic anions accumulated by the lupins was not different ($P < 0.05$) between the 250 and 5000 $\mu$M NO$_3^-$ treatments, with an average rate of 872 $\mu$eq g$^{-1}$ d$^{-1}$ (Table 2). About 82% of the organic anions accumulated in the roots of lupins for both N treatments. Peas accumulated organic acids at rates of 1235 and 395 $\mu$eq g$^{-1}$ d$^{-1}$, when supplied with 250 and 5000 $\mu$M NO$_3^-$ respectively. The proportion of the organic anions accumulated in the pea shoots was 16 and 60% when supplied with 250 and 5000 $\mu$M NO$_3^-$, respectively.

The lupins accumulated mainly malate in the shoots and citrate in the roots, and there was no effect ($P < 0.05$) of the treatments on these proportions (Fig. 2). Similarly, peas accumulated mainly malate in their shoots and citrate in their roots. However more malate was present when the peas were supplied with 5000 $\mu$M than 250 $\mu$M NO$_3^-$ in both their shoots and roots.

Calculated organic anion excretion. The calculated excretion from lupins supplied with 250 and 750 $\mu$M NO$_3^-$ was negligible (less than 21 $\mu$eq g$^{-1}$ d$^{-1}$; Table 2). Using eqn (2), it was calculated that lupins excreted organic anions at rates of 189 and 770 $\mu$eq g$^{-1}$ d$^{-1}$ when supplied with 2500 and 5000 $\mu$M NO$_3^-$ respectively.

Peas excreted less than 101 $\mu$eq g$^{-1}$ d$^{-1}$ but because of the large standard errors, particularly with the $H^+$ excretion measurement, these quantities were not significant ($P < 0.05$).

Experiment 2

The growth of the lupins during the experiments was satisfactory, and no signs of nutrient deficiencies were evident. There was no effect ($P < 0.01$) of the treatments on the root or shoot growth during the expt 2 (Table 3).

Sterile conditions were only maintained for about the first 3 weeks of the pretreatment period, after which contamination entered the culture system. Although contaminated, the decomposition of the organic anions was much less than in a preliminary experiment. An average of 42 $\mu$g ml$^{-1}$ of the 100 $\mu$g ml$^{-1}$ citrate added to the vials was recovered after the 4 d treatment periods, a decomposition rate of 0.6 $\mu$g ml$^{-1}$ h$^{-1}$. This was about one tenth of the decomposition rate measured in a preliminary experiment where lupins were grown in a non-sterile culture system, similar to expt 1.

No significant ($P < 0.05$) quantities of organic acids were detected after the first 4 d collection period and only results from the second collection period are presented and discussed.

Cation–anion balance and $H^+$/OH$^-$ excretion. The uptake of nutrients in expt 2 was at similar rates to those in expt 1. There was no difference ($P < 0.05$) between the excess cation uptake and $H^+$ excreted by lupin roots which were not supplied with NO$_3^-$ (Table 4). When supplied with NO$_3^-$, lupins absorbed more Ca$^{2+}$ and K$^+$ and less SO$_4^{2-}$ than when not supplied with NO$_3^-$; however, the uptake of NO$_3^-$ counteracted these effects and an excess of anion uptake resulted. Despite the excess of anion uptake, the lupins
Organic acid anion accumulation. Unlike exp 1, there was a trend for the lupins to accumulate less organic anions with increasing NO₃ supply (Table 4). The proportions and types of organic anions accumulated in the roots and shoots were not affected by the NO₃ supply. An average of 86% of the organic anions in the plant were in the roots, of which 81% was citrate.

Organic acid anion excretion. Of the organic anions excreted, 98% was citrate with the remainder being malate. Lupins excreted up to 87 μeq g⁻¹ d⁻¹ organic anions and there was a strong trend of increasing organic anion excretion with increasing excess of anion uptake and NO₃ uptake (Table 4). Despite the microbial contamination, there was close agreement between the organic anion excretion calculated from the model and that estimated by direct measurements ($r^2 = 0.85$).

DISCUSSION

Lupins and peas have different mechanisms of maintaining internal charge balance when supplied with NO₃ and a different metabolism of organic acid anions. Lupins absorbed less NO₃ than peas on a dry weight basis at the same NO₃ supply, which largely accounted for the smaller excess of inorganic anion uptake over inorganic cation uptake in lupins than in peas. When inorganic anion uptake exceeded inorganic cation uptake, lupins excreted an equivalent negative charge as citrate and a small amount of OH⁻. In exp 1, peas excreted OH⁻ in amounts equivalent to their excess of inorganic anion over inorganic cation uptake, and no significant quantity of organic anion excretion was calculated.

The majority of the organic anions accumulated in the roots of the lupin and in the shoots of peas, which relates to the proportion of NO₃ reduction in the roots and shoots for both species when supplied with moderate concentrations of NO₃ (Andrews et al., 1984). Sulphate reduction is widely spread throughout the organs of higher plants (Schiff, 1983), and is proportionally smaller than NO₃ reduction. Hence, SO₄²⁻ reduction may have little effect on the distribution of organic anion accumulation. Van Beusichem et al. (1985) showed that 80% of the NO₃ and SO₄²⁻ reduction and a large proportion of organic anion accumulation was located in the shoots of castor oil plants.

Why lupins should excrete citrate rather than OH⁻ is not clear. Organic anions may increase the availability of some nutrients. Gardner, Parbery and Barber (1982) and Dinke-laker, Römheld and Marschner (1989) measured exudates from the proteoid roots of P and Fe deficient white lupins (L. albus L.) in soil. They noted that the exudates were ‘neutral in pH’ and contained large quantities of citrate. While the narrow-leaved lupins used in this study do not form proteoid roots and did not show signs of P or Fe deficiency, organic anion excretion occurred. Unlike peas, the root growth of L. angustifolius L. is decreased by increasing the solution pH from 5.5 to 6.0 (Tang et al., 1992), and the excretion of organic anions rather than OH⁻ when inorganic anion uptake exceeds inorganic cation...
uptake is perhaps a method of preventing high pH in the rhizosphere.

The excretion of citrate by lupins may explain their relatively high acid and aluminium tolerance, because citrate is a strong chelator capable of detoxifying Al³⁺. Christiansen-Weniger, Gronemen and Van Veen (1992) demonstrated that the aluminium tolerance of wheat cultivars was closely related to the amount of organic anion excretion from their roots.

Growing seedling roots in sterile conditions was not achieved despite the measures taken in this study. Kerven et al. (1991) also did not achieve complete sterility when plants were grown in 30 mg l⁻¹ cefotaxime, however they did not measure any significant loss of organic acids from their system due to microbial decomposition. It might be expected that the micro-organisms will break down the organic anions to OH⁻ and CO₂, however there was no increase in the pH of the nutrient solution in these and other experiments (Atwell, 1992; Loss et al., 1993b). Micro-organisms may convert the organic anions into microbial biomass containing carboxylate groups that do not affect the pH of the external nutrient solution. It is not known if this would also occur in soil.

These results have implications for subsoil acidification under lupins in the mediterranean-type climate of Western Australia and elsewhere. The cultivation of lupins for grain (L. angustifolius L.) causes increased soil acidification, particularly in the subsoil where more shallow rooted legumes have little effect on soil pH (Coventry and Slattery, 1991; Loss, Ritchie and Robson, 1993a). In a previous study (Loss et al., 1993b), nodulated lupins were grown in a vertical split pot which allowed the upper and lower zones of roots to be supplied with varying concentrations of K⁺ and NO₃⁻. Proton excretion was not distributed evenly over the entire root length but was concentrated in zones of high cation uptake, and hence differences in nutrient uptake by roots between the surface soil and the subsoil will lead to different rates of H⁺ excretion.

Nitrate is rapidly leached in the coarse sandy soils to which lupins are best adapted in Western Australia (Diggle, Bowden and D’Antuono, 1990) and up to 10 mm NO₃⁻ has been measured in the soil solution extracted from a sandy subsoil (Carr, pers. comm.). Although lupins are reputed to have a low capacity to absorb NO₃⁻, they may absorb significant quantities from the subsoil with the onset of the summer drought when the soil nearer the surface begins to dry. Previous results (Loss et al., 1993b) indicate that plants maintain their electroneutrality at the site of nutrient uptake and the excess anion uptake in the subsoil roots would result in the excretion of citrate in the subsoil. Provided the pH of the soil is greater than 5.4, the pKa₂ value of citrate, most of the excreted citrate would remain as an anion and would not affect the soil pH. Combined with cation uptake, particularly K⁺, this mechanism could lead to considerably more subsoil acidification under lupins than under other plant species, as was measured in the field (Loss et al., 1993a).

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LITERATURE CITED


