Light scattering determination of magnetic moments of magnetotactic bacteria (invited)

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Light scattering is used to determine the average lengths and magnetic moments of magnetotactic bacteria in culture. The results are consistent with estimates made from electron micrographs.

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INTRODUCTION

Magnetotactic bacteria from aquatic sediments orient and swim along magnetic field lines.1 Each cell contains single magnetic domain Fe3O4 particles that impart a magnetic dipole moment to the cell, parallel to the axis of motility.2,3 The cells migrate in the field direction (North-seeking) or opposite to the axis of motility.2,3 The cells migrate in the field direction (South-seeking) depending on the orientation of the moment in the cell. The vertical component of the geomagnetic field selects the predominant cell polarity in natural environments.4,5

In Aquaspirillum magnetotacticum, a freshwater spirillum grown in pure cultureq theoFe304 particles are typically cuboidal of 400 - 500 A dimension.7 The average number of particles per bacterium can vary from zero to as many as forty depending on the conditions and the concentration of nutrients and dissolved oxygen.

Light scattering was used to determine both the average length and magnetic moment of the bacteria in cultures SL and S2. Since the volume of Fe3O4 is only about 1/1000 of the volume of a bacterium, we assume the scattering strength is uniform over the entire bacterium. Thus, modeling the bacteria as optically isotropic cylinders of length L and radius R, one can show that the single particle static structure factor is given by

\[ S_{\text{sp}}(Q_L, Q_R, \phi) = \frac{\sin(Q_L \cos \phi)}{Q_L^2 \cos \phi} \left( \frac{2 J_1(Q_R \sin \phi)}{Q_R \sin \phi} \right)^2 \] (1)

where \( J_1 \) is the Bessel function of order 1 and \( \phi \) is the angle between \( Q \) and the long axis of the bacterium. \( Q \) is the momentum transfer of light (\( Q = k_L - k_f \)). For small diameter bacteria, \( Q \ll 1 \) and \( \sin \phi \approx \phi \) and thus the corrections caused by the spiral nature of the bacterium to a small rescaling of L. The assumption of orientational optical isotropy is valid because it was determined that the birefringence of a fully aligned sample in vitro is about 10^-4, and thus should cause only insignificant depolarization effects.

If a magnetic field \( \vec{H} \) is applied parallel to \( \vec{Q} \), bacteria of magnetic moment \( \mu \) orient parallel to both \( \vec{Q} \) and \( \vec{H} \) with an angular distribution function \( P(\phi, \alpha) \) given by

\[ P(\phi, \alpha) = \frac{a \cos \phi \sin \alpha}{2 \sinh a} \] (2)

where \( a = \mu H / k_B T \), \( k_B \) is Boltzmann's constant, \( T \) is temperature, and \( \alpha \) is defined as before. The total structure factor \( S(Q_L, Q_R, \alpha) \) is therefore the single particle structure factor, weighted by the angular distribution function, integrated over all angles:

\[ S(Q_L, Q_R, \alpha) = N \int P(\phi, \alpha) S_{\text{sp}}(Q_L, Q_R, \phi) \, d\phi \] (3)

where \( N \) is the number of scatterers and \( Q_L \) is parallel to \( \vec{H} \). The scattered light intensity is thus proportional to \( S(Q_L, Q_R, \alpha) \).

Three Helmholtz coil pairs oriented at right angles to each other were used to cancel the ambient magnetic field to less than 0.01 Oe at the sample position. A fourth Helmholtz coil pair produced magnetic fields from 0 to 40 Oe parallel to \( \vec{Q} \) at the sample.

To perform the experiment, the wavevector \( Q \) was chosen to be 9998 cm^-1, with an acceptance width of 500 cm^-1 (Fig. 2). The average scattering intensities, measured with a phototube in conjunction with a low pass filter (time constant \( t = 13.5 \) sec), were cor-
Theoretical plot of $R(Q_{L},a)$ vs $a$ for a particular value of $Q_{L}(Q_{L} = 3.48)$ where $R(Q_{L},a) = S(Q_{L},a)/S(Q_{L},0)$. For a given applied field, $R(Q_{L},a)$ was measured, thus establishing $a_{0} = k_{B}T/H$ was obtained. In zero field the alignment is random and in high field ($a > 100$) it is saturated, with all bacteria aligned parallel to $H$. $<\lambda>$ was obtained by measuring $S(QL,a)$ in zero field and in a saturation field of 40 Oe, for which it was later determined that $a = 200$. The quantity $R(QL,a) = S(QL,a)/S(QL,0)$, which rapidly approaches a limiting value for $a > 100$, is a strong function of $Q_{L}$, as can be seen in Fig. 3 by comparing the curves for $a = 0$ and $a = 100$. Thus a measurement of the intensity ratio $R(QL,a)$ for $a > 100$ accurately determines $Q_{L}$, and since $Q$ is known from the scattering geometry, $<\lambda>$ is determined. In Fig. 3 we show the measured value of $QL$ for culture S2. Values of $<\lambda>$ for S1 and S2 are given in Table I.

Having determined $QL$, and therefore $<\lambda>$, using the saturation field, $<\lambda>$ is obtained by measuring $R(QL,a)$ at many different fields over the range zero to 40 Oe. In Fig. 4, $R(QL = 3.48,a)$, corresponding to the dashed line in Fig. 3, is shown. For a given field $H$, the value of $a$ corresponding to the measured value of $R(QL,a)$ is determined from Fig. 4. Since $a = uH/k_{B}T$, and $H$ and $T$ are known, $<\lambda>$ is obtained. This measurement was performed for approximately ten values of $H$ for each culture and results were averaged to obtain $<\lambda>$. Values of $<\lambda>$ are listed in Table I. We note in passing, however, that since $R(QL,a)$ is a strong function of $a$, the distribution width $\sigma_{a}/<\lambda>$ must be less than 0.2 to obtain an accurate measurement of $<\lambda>$.

In addition to the light scattering measurement, a birefringence technique, similar to that of Scholten, was used to measure $<\lambda>$. Here the bacteria are assumed to possess an optical polarizability anisotropy $\delta_{p}$, so that the sample birefringence is

$$\Delta n = N\delta_{p} <\lambda_{1}/2 - 1/2>$$  \hspace{1cm} (4)

where $N$ is the number of bacteria and $\delta_{p}$ is the angle between $H$ and the long axis of the bacterium. This technique is insensitive to $L$ and preliminary results for $<\lambda>$ were consistent with those of the light scattering method. Details will be published elsewhere.
TABLE I. Parameters of Magnetotactic Bacteria Determined by Light Scattering (L.S.) and Electron Microscopy (E.M.)

<table>
<thead>
<tr>
<th></th>
<th>Sample 1</th>
<th>Sample 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Average Length (Microns)</td>
<td>2.93</td>
<td>3.52</td>
</tr>
<tr>
<td>E.M.</td>
<td>3.2 ± 0.6</td>
<td>3.4 ± 0.8</td>
</tr>
<tr>
<td>Average Magnetic Moment (emu)</td>
<td>(2.2±0.2)x10^{-13}</td>
<td>(4.3±0.5)x10^{-13}</td>
</tr>
<tr>
<td>E.M.</td>
<td>2.7±0.5)x10^{-13}</td>
<td>(5.0±0.8)x10^{-13}</td>
</tr>
</tbody>
</table>

CONCLUSIONS

Considering the uncertainties in estimating the magnetic moments from the electron micrographs, the agreement of the moments obtained by light scattering with those estimates is quite good. Especially important is the fact that both methods give the same ratio for the average moments in the two samples. Thus static light scattering is a reliable method for rapidly determining the average lengths and magnetic moments of magnetotactic bacteria in culture, and can be used to assay these physical characteristics for bacteria grown under differing culture conditions.

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REFERENCES