VINYL CHLORIDE BIODEGRADATION WITH METHANOTROPHIC ATTACHED FILMS

By Yarrow M. Nelson, and William J. Jewell

ABSTRACT: Methanotrophic degradation of vinyl chloride (VC) is investigated using a laboratory-scale methanotrophic attached-film expanded-bed (MAFEB) bioreactor. This study provides a basis for applying a microbial cometabolizing reaction to practical treatment of toxic chlorinated compounds. The MAFEB reactor was operated at 20°C with influent VC concentrations ranging from 1,800 to 9,600 μg/L and bed hydraulic retention times ranging from 3.7 to 7.6 h. VC effluent concentrations during steady continuous operation ranged from 3 to 140 μg/L, with most values less than 26 μg/L, resulting in removal efficiencies of 96.3% to 99.8%. The maximum continuous-flow VC degradation rate observed at 20°C was 2.5 mg VC per gram volatile solids (VS) per day [2.5 mg VC/(g VS d)] or 30 mg VC per liter expanded bed per day [30 mg VC/(Lreh d)], under substrate-limited conditions. During semibatch runs at 35°C, vinyl chloride degradation rates up to 60 mg VC/(g VS d) or 1 g/(Lreh d) were observed. Degradation rates increased with temperature between 20°C and 35°C, approximately doubling every 10°C. Dissolved methane concentrations above 0.5 mg/L inhibited VC degradation, with no VC degradation observed with 8 mg/L dissolved methane. The methane consumed during VC degradation was about 40 g CH4/g VC. Toxic effects were observed after prolonged exposure of the methanotrophic culture to high concentrations of VC.

INTRODUCTION

Chlorinated ethenes, such as tetrachloroethene (PCE), trichloroethene (TCE), dichloroethene (DCE), and chloroethene [vinyl chloride (VC)] are toxic and suspected carcinogens (Craun 1984) and are among the most common ground-water pollutants (Westrick et al. 1984; Love et al. 1982). Biodegradation of TCE, the most common of these compounds, has been studied extensively, while relatively few studies have been conducted on the biodegradation of VC (Chaudhry et al. 1991). Although VC is not expected to enter an aquifer directly, it has been detected at sites contaminated with PCE, TCE, and DCE, presumably as a result of anaerobic reductive dechlorination of these compounds (Parsons et al. 1984, 1985; Milde 1988; Molton 1987). This production of VC is of concern because of the high toxicity and known carcinogenicity of VC (“Ambient” 1980; Malti 1977; Lee et al. 1978; Infante 1982). In situ anaerobic remediation processes can also lead to the formation of VC (Vogel and McCarty, 1985).

Vinyl chloride formation must also be considered in the design of aboveground treatment schemes for remediation of contaminated ground-water sites. Ideally, an aboveground anaerobic treatment system for PCE or TCE removal would dechlorinate these compounds to ethene and/or carbon dioxide and chloride ions. However, anaerobic systems have been reported to result in the accumulation of VC as an intermediate (Vogel et al. 1987), particularly when operating at typical ambient groundwater temperatures.
The presence of VC can also cause problems for aboveground remediation with physical processes. The release of VC during air stripping is becoming unacceptable, and adsorption of VC by activated carbon is limited compared to that of less soluble chlorinated compounds such as PCE and TCE.

We have proposed a two-stage treatment system for complete biodegradation of a range of chlorinated compounds, consisting of an anaerobic first stage followed by an aerobic methanotrophic second stage. The anaerobic stage is expected to dechlorinate highly chlorinated compounds, such as PCE and TCE, while the methanotrophic stage would biodegrade the dechlorination products from the first stage. This hybrid system would utilize the best capabilities of both microbial populations, because anaerobic cultures have been found to degrade highly chlorinated compounds more rapidly than less chlorinated compounds (Vogel et al. 1987; Freedman and Gossett 1989), while methanotrophic cultures have been found to degrade less-chlorinated compounds more rapidly than highly chlorinated compounds (Fogel et al. 1986; Tsien et al. 1989). In fact, methanotrophic degradation of PCE has been reported to be minimal or nonexistent (Fogel et al. 1986; Tsien et al. 1989), and PCE has also been shown to be toxic to methanotrophs (Fennell et al. 1992). Sequential anaerobic/aerobic biodegradation of PCE and TCE has been proposed previously (Fathepure et al. 1987), and the feasibility of applying such a system to in situ remediation has been investigated in a simulated aquifer (Dooley-Danna et al. 1989; Fogel 1990).

Biodegradation of VC is essential to efficient operation of the two-stage system because of the significant quantities of VC expected to be produced by the anaerobic stage, and because VC is considered more toxic and carcinogenic than TCE or PCE. Anaerobic cultures have been found to degrade VC by a number of mechanisms (Barrio-Lage et al. 1990; Vogel et al. 1987), but such degradation is slow and inefficient, especially at reduced temperatures (Freedman and Gossett 1989; Chu 1991; Carter 1991). Hartmans et al. (1985) reported that Mycobacterium L1, an aerobic organism, was capable of growing on VC as the sole carbon and energy source. More recently, Davis et al. (1990) reported on aerobic degradation of VC by naturally occurring microorganisms in simulated aquifers. Several studies have briefly examined VC degradation by methanotrophic bacteria (Fogel et al. 1986, 1987; Tsien et al. 1989; Roberts et al. 1989). A VC degradation rate of 14.6 nmol/(mg h) [22 mg/(g d)] was reported by Fogel et al. (1987) using a methanotrophic consortium (temperature not specified). Detailed kinetics of methanotrophic VC degradation remain to be defined, especially for continuous-flow operation at temperatures characteristic of ground water. This work defines the kinetics and feasibility of methanotrophic VC biodegradation in a continuous-flow laboratory-scale reactor.

Experiments were conducted in a methanotrophic attached-film expanded-bed (MAFEB) reactor, which has been described previously (Fennell and Jewell 1992). The primary advantage of an attached-film reactor for this application is the high solids concentration, making operation with short hydraulic retention times possible. It was also hypothesized that the biofilms would enhance the cometabolism reactions, since the primary electron source (dissolved methane) could inhibit VC degradation. Another advantage of this type of reactor is the high effluent quality achieved without secondary settling.

To define the kinetics of VC degradation, a series of continuous-flow and
semibatch runs were conducted in laboratory-scale MAFEB reactors. Semibatch runs with the same bioreactor were conducted to evaluate the effects of temperature and methane concentration. Toxicity of VC to the methanotrophic culture was also investigated during continuous-flow operation. This work was part of a larger study that has been ongoing since 1988 (Jewell et al. 1990) sponsored by the Gas Research Institute and the U.S. Department of Energy to define processes for aboveground and in situ treatment of contaminated ground waters.

EXPERIMENTAL

MAFEB Reactor Design

The MAFEB reactor system (Fig. 1) was similar to MAFEB systems described by Fennell et al. (1992). The 5 cm diameter expanded-bed volume was 0.4 to 0.8 L, and the total liquid volume in the system was 3.8 L. A 10

FIG. 1. Schematic of MAFEB Reactor System Used for Methanotrophic VC Degradation Experiments
to 20 cm clarifier zone was employed above the bed to allow settling of suspended solids. Liquid recycled above the bed was divided into two separate bubble diffusers where oxygen and methane gas were introduced. The gas exchangers were 100 cm tall and 3.8 cm in diameter, expanding to 5 cm at the bottom to accommodate two diffuser stones. Gases were continuously recycled through both diffusers. Make-up oxygen and methane (Matheson ultra high purity) were supplied to each diffuser from compressed gas cylinders via low-pressure regulators set at 3 kPa (30 cm water). Gas was fed from 6.9 L pressurized tanks that were refilled as needed with oxygen or methane. Gas-use rates were monitored by recording the pressure in these supply cylinders. The reactor system was constructed primarily with glass, stainless steel, polyvinyl chloride, and Teflon components because these materials were found to have minimal reactivity with VC. The reactor system was housed in a temperature-controlled chamber.

**Reactor Operation**

Fused diatomaceous earth particles between 300 and 1,000 μm in diameter were used as support media for the methanotrophic films in the expanded bed (Eagle-Picher Minerals, Inc., Cincinnati). Particles with mature methanotrophic films were obtained from a larger MAPEB reactor that was operated as a breeder reactor. This breeder reactor was initially inoculated with settled sewage and soil samples from a natural gas outgassing site. Nutrients were supplied in excess either from secondary sewage effluent or from synthetic nutrient mixtures (Table 1). The mixtures consisted of NH₄Cl, equimolar amounts of K₂HPO₄ and KH₂PO₄, and yeast extract (BBL Microbiology Systems). Secondary sewage effluent was obtained from a local wastewater-treatment facility.

The reactor was operated with liquid recycle rates of 250 mL/min from each diffuser, making the hydraulic retention time in the bed about one minute per pass. The gas recycle rates were 150 mL/min and 100 mL/min for the oxygen and methane diffusers, respectively. Gas recycle rates were adjusted periodically to control the composition of gases in the diffuser headspaces and to control the dissolved methane concentrations in the reactor. Dissolved gas (CH₄ and O₂) concentrations were measured using a headspace analysis technique with liquid samples. Since copper has been reported to affect methanotrophic yields and MMO enzyme activity (Leak and Dalton 1986; Brusseau et al. 1990), the copper-ion concentrations in the reactor were monitored using inductively coupled plasma spectroscopy.

**TABLE 1. Summary of Operating Parameter during VC Continuous-Flow Runs**

<table>
<thead>
<tr>
<th>Condition (1)</th>
<th>1 (2)</th>
<th>2 (3)</th>
<th>3 (4)</th>
<th>4 (5)</th>
<th>5 (6)</th>
<th>6 (7)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Feed rate (L/day)</td>
<td>1.5-3.0</td>
<td>3.0</td>
<td>1.9</td>
<td>2.0</td>
<td>2.6</td>
<td>0-3</td>
</tr>
<tr>
<td>Expanded-bed volume (L)</td>
<td>0.46</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
<td>0.6</td>
<td>0.5</td>
</tr>
<tr>
<td>HRT in expanded bed (h)</td>
<td>3.7-7.4</td>
<td>4.8</td>
<td>7.6</td>
<td>7.2</td>
<td>5.5</td>
<td>4.0</td>
</tr>
<tr>
<td>Influent VC concentration (mg/L)</td>
<td>8.9</td>
<td>2.9</td>
<td>3.0</td>
<td>1.5</td>
<td>2.0</td>
<td>4-8</td>
</tr>
<tr>
<td>VC loading rate (mg VC/L&lt;sub&gt;0&lt;/sub&gt; d)</td>
<td>29-58</td>
<td>14.5</td>
<td>9.5</td>
<td>5.0</td>
<td>8.7</td>
<td>0-48</td>
</tr>
<tr>
<td>Nutrient concentrations in Feed</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Source of feed</td>
<td>Synthetic</td>
<td>Synthetic</td>
<td>Sewage</td>
<td>Sewage</td>
<td>Sewage</td>
<td>Synthetic</td>
</tr>
<tr>
<td>Ammonia (mg N/L)</td>
<td>44</td>
<td>19</td>
<td>14</td>
<td>18</td>
<td>17</td>
<td>40</td>
</tr>
<tr>
<td>Phosphate (mg P/L)</td>
<td>3.4</td>
<td>2.8</td>
<td>1.0</td>
<td>1.1</td>
<td>0.9</td>
<td>4</td>
</tr>
<tr>
<td>Yeast extract (mg/L)</td>
<td>5</td>
<td>2.5</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2.5</td>
</tr>
<tr>
<td>Duration of experiment (days)</td>
<td>22</td>
<td>15</td>
<td>8</td>
<td>8</td>
<td>7</td>
<td>27</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>20</td>
<td>20</td>
<td>20</td>
<td>20</td>
<td>20</td>
<td>19</td>
</tr>
</tbody>
</table>
Continuous-Flow Runs

Six continuous-flow runs with VC were operated at 20°C ± 1°C (Table 1). VC was analyzed daily by headspace gas chromatographic analysis of influent and effluent liquid samples. Influent samples were collected from the feed line upstream from the feed pump, and effluent samples were collected from a sample port in the liquid recycle line. Dissolved methane and oxygen were measured by a similar technique for samples collected above and below the expanded bed. Feed rates were varied in response to observed VC toxicity effects. As methane inhibition became evident, the gas recycle rates were adjusted to control the dissolved methane concentration in the reactor.

Semibatch Runs

Semibatch experiments were conducted with the same apparatus and attached film biomass as used for the continuous-flow runs. VC was added to the reactor system at the beginning of each run, while oxygen and methane were supplied continuously. At the beginning of the tests, pure VC gas (Alphagaz Specialty Gas Products) was injected into the methane-gas recycle line. Just prior to injecting VC to start the semibatch runs, 100 mL of nutrient stock solution (800 mgN/L and 80 mgP/L) was pumped into the reactor. Liquid samples were taken from the liquid recycle line below the expanded bed (Fig. 1), starting about 5 min after VC injection.

Semibatch runs for the methane inhibition study were conducted at 20°C and operated with a pH between 5 and 6. These runs were initiated with 5 mL of VC gas that resulted in initial aqueous VC concentrations of about 1.5 mg/L. The expanded-bed volume varied from 0.35 to 0.40 L, and the total biomass content of the reactor varied from 6.2 to 8.2 g volatile solids (VS). Total biomass content at the time of each run was estimated from initial and final measurements and from the amount of methane used up to the time the semibatch run was initiated. Gas recycle rates were adjusted to control the dissolved methane concentration in the reactor.

Temperature effects were examined between 20°C and 35°C. These runs were initiated with 15 mL of VC gas injected into the reactor, which resulted in initial VC concentrations of 3 to 5 mg/L. Reactor effluent pH was similar for all of these semibatch runs (5.3 to 5.5). Prior to and in between each of these semibatch runs, the MAFEB reactor was operated in continuous-flow mode without VC to maintain growth of the methanotrophic culture. The methanotrophic culture was allowed to acclimate to each new temperature by operating the reactor in continuous-flow mode for one to two weeks (10 to 15 total system hydraulic retention times) prior to each semibatch run.

Reactor Control Tests

The integrity of the reactor system with respect to VC loss was evaluated using semibatch and continuous-flow reactor control tests. These tests were conducted with a clean reactor system that was filled with distilled water and purged with nitrogen. Semibatch reactor control tests were initiated by injecting 15 mL of VC gas via the methane gas recycle line, which resulted in an initial aqueous VC concentration of 6.5 mg/L. During the semibatch control runs, all recycle pumps were operated at the same rates as in actual semibatch runs. After 100 h of operation, the VC concentration measured in the reactor remained within 0.5 mg/L of the initial value. Continuous-flow reactor control tests were operated by flowing a 10 mg/L solution of
VC through the reactor system at a rate of 3 L/d. Liquid samples were collected from the reactor influent and effluent ports twice daily for analysis. This test resulted in less than 6% variation between influent and effluent VC concentrations for six days of operation.

The semibatch and continuous-flow control tests confirmed that the reactor system itself does not adsorb or leak VC. Adsorption of VC by the bed material was investigated during a semibatch run in which biodegradation of VC was inhibited with high dissolved methane concentrations. Under these conditions, little or no loss of VC was observed, indicating that VC was not lost due to adsorption by the bed. These results rule out any short-term VC adsorption or loss during the semibatch tests. It can be concluded that all reductions of VC during the experiments in the MAFEB reactor are attributable only to biodegradation.

**Analytical**

VC concentrations in gas samples from the MAFEB reactor headspaces were determined by direct gas chromatography of the samples. VC concentrations in liquid samples were determined by analyzing the gas headspace of samples in 5 mL reaction vials (Supelco, Inc., Bellefonte, Pa.). VC in the headspace of these samples was analyzed with an HP 5890 gas chromatograph (GC) (Hewlett Packard) equipped with a flame ionization detector and an 2.4 m (8 ft) long, 3.9 mm (1/8 in.) inside diameter (ID) stainless-steel column packed with SP-1000 on 60/80 Carbopak B (Supelco). Helium was used as the carrier gas at a flow rate of 35 mL/min and a pressure of 400 kPa (60 psi). The column temperature was 160°C, which resulted in VC retention times of 0.65 min. The total amount of VC in the sample was determined using Henry’s constants published by Gossett (1987).

Gas concentrations of O₂, CH₄, CO₂, and N₂ were determined by gas chromatography using a Gow Mac Series 580 GC with a thermal-conductivity detector. The chromatograph was equipped with two 1.7 m steel columns (0.6 cm ID). The columns were packed with 60/80 mesh Poropak Q (Supelco) for CH₄ and CO₂ measurement and with 60/80 mesh Molecular Sieve 5A (Supelco) for O₂ and N₂ measurement.

Dissolved methane concentrations were measured using a headspace GC technique. Headspaces of sealed liquid samples were analyzed for methane using the Gow Mac GC equipped as described previously. Liquid samples were collected with a glass syringe from the recycle line just below the expanded bed, or from an effluent port above the expanded bed. Since the retention time of liquid in the bed per pass was usually only 1 min, the reactor approached completely mixed conditions. However, rapid uptake of methane in the bed caused a differential between the influent and effluent dissolved methane concentration averaging about 0.5 mg/L.

Total suspended and volatile suspended solids of the reactor liquid were determined by methods 209C and 209D, respectively, of *Standard Methods*, 16th Ed. (1985). This procedure was also used to analyze solids removed from the reactor during cleaning to assay the volatile solids content of the wall growth and sludge in the reactor. Volatile solids attached and entrapped in the expanded bed were determined by a technique described by Clarkson (1986), in which samples of the bed were weighed after drying and after ashing at 550°C.
RESULTS

General Reactor Operation

Well-defined operation of the methanotrophic attached-film expanded-bed reactor was sustained for a period of about 2 years, during which time these experiments with vinyl chloride were conducted. The MAFEB system operated with similar growth characteristics and gas-use rates as those previously observed for this type of reactor system, as shown in Table 2 and Fennell and Jewell (1992). The total quantity of attached film was limited to one third to one 10th of the maximum potential, which appeared to be around 50 g VS/L in earlier long-term tests (Fennell and Jewell 1992). Growth rates and yields varied significantly between runs due to differences in operating conditions. Observed yields were similar to those reported for methanotrophs in the literature (Vary and Johnson 1967), except during runs CF-1 and CF-6 during which high VC concentrations may have limited growth. Another factor affecting growth rates was the rate of methane gas recyle, which was reduced during runs CF-4 through CF-6 to avoid suspected methane inhibition of VC degradation.

Growth on the walls of the reactor and suspended in the diffusers accounted for 10% to 20% of the total growth, and the remainder of the growth occurred in the expanded bed. The reactor was operated without addition of alkalinity to the feed, and therefore CO2 evolved by the methanotrophs resulted in a low operating pH of between 5.5 and 6.0 (Table 2). In contrast to studies that indicate an optimum pH for growth near neutral (Vary and Johnson 1967), our cultures appeared to grow more rapidly at a pH near 6.0.

Continuous-Flow VC Degradation

The MAFEB reactor was operated for six continuous-flow runs with influent VC concentrations ranging from 1,000 to 10,000 µg/L and expanded-bed hydraulic retention times ranging from 3.7 to 7.6 h (Table 1). The continuous-flow runs with relatively low VC loading rates (2–5) proceeded with rapid and consistent VC degradation (Fig. 2). Average VC removal

<table>
<thead>
<tr>
<th>Condition</th>
<th>Run Number 1 (2)</th>
<th>2 (3)</th>
<th>3 (4)</th>
<th>4 (5)</th>
<th>5 (6)</th>
<th>6 (7)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Initial Seed (g VS)</td>
<td>2.7</td>
<td>3.5</td>
<td>2.0</td>
<td>3.7</td>
<td>2.9</td>
<td>3.1</td>
</tr>
<tr>
<td>Final Biomass:</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bed Conc., attached (g VS/L)b</td>
<td>9.3</td>
<td>14</td>
<td>7.6</td>
<td>5.8</td>
<td>5.8</td>
<td>5.2</td>
</tr>
<tr>
<td>Bed, attached (g VS)</td>
<td>4.3</td>
<td>8.6</td>
<td>4.6</td>
<td>3.5</td>
<td>3.5</td>
<td>2.6</td>
</tr>
<tr>
<td>Bed, Entrapped (g VS)</td>
<td>1.4</td>
<td>0.79</td>
<td>0.55</td>
<td>0.83</td>
<td>0.35</td>
<td>0.20</td>
</tr>
<tr>
<td>Wall Growth/Sludge (g VS)</td>
<td>0.66</td>
<td>0.67</td>
<td>0.88</td>
<td>1.10</td>
<td>0.27</td>
<td>0.86</td>
</tr>
<tr>
<td>Average Biomass in System (g VS)</td>
<td>5.8</td>
<td>6.7</td>
<td>4.0</td>
<td>4.5</td>
<td>4.1</td>
<td>3.4</td>
</tr>
<tr>
<td>Suspended Solids (mg VSS/L)</td>
<td>13</td>
<td>11</td>
<td>30</td>
<td>30</td>
<td>7</td>
<td>7</td>
</tr>
<tr>
<td>Total SS Wasted to Effluent (g VSS)</td>
<td>0.46</td>
<td>0.33</td>
<td>0.53</td>
<td>0.43</td>
<td>0.11</td>
<td>0.30</td>
</tr>
<tr>
<td>Daily Growth Rate (g VS/d)</td>
<td>0.05</td>
<td>0.62</td>
<td>0.66</td>
<td>0.30</td>
<td>0.22</td>
<td>0.04</td>
</tr>
<tr>
<td>Avg. Observed Yield (g VS/g COD)</td>
<td>0.03</td>
<td>0.13</td>
<td>0.15</td>
<td>0.07</td>
<td>0.08</td>
<td>0.03</td>
</tr>
<tr>
<td>Methane use average (L/d)</td>
<td>0.59</td>
<td>1.7</td>
<td>1.6</td>
<td>1.2</td>
<td>0.9</td>
<td>0.4</td>
</tr>
<tr>
<td>Oxygen use average (L/d)</td>
<td>0.69</td>
<td>2.5</td>
<td>2.5</td>
<td>1.9</td>
<td>1.6</td>
<td>0.9</td>
</tr>
<tr>
<td>Oxygen/Methane Ratio (L/L)</td>
<td>1.2</td>
<td>1.5</td>
<td>1.6</td>
<td>1.6</td>
<td>1.7</td>
<td>1.5</td>
</tr>
<tr>
<td>Avg. Effluent pH</td>
<td>5.5–6.8</td>
<td>5.5</td>
<td>5.8</td>
<td>5.9</td>
<td>6.1</td>
<td>5.8</td>
</tr>
</tbody>
</table>
FIG. 2. Influent and Effluent VC Concentrations during Continuous-Flow Runs in the MAFEB Reactor without Observed VC Toxicity
TABLE 3. Summary of Vinyl Degradation Steady Parameter of VC Continuous-Flow Runs

<table>
<thead>
<tr>
<th>Condition</th>
<th>Run Number</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1(2)</td>
</tr>
<tr>
<td>Average influent VC concentration (μg/L) b</td>
<td>8,900(900)</td>
</tr>
<tr>
<td>Average effluent VC concentration (μg/L) b</td>
<td>26(8)</td>
</tr>
<tr>
<td>Removal Efficiency (%)</td>
<td>99.7</td>
</tr>
<tr>
<td>Average dissolved methane concentration (mg/L) c</td>
<td>0.2</td>
</tr>
<tr>
<td>VC Degradation rates:</td>
<td></td>
</tr>
<tr>
<td>Biomass basis (mg VC/g VS-day)</td>
<td>1.7</td>
</tr>
<tr>
<td>Exp. Bed basis (mg VC/L eb-day)</td>
<td>20</td>
</tr>
<tr>
<td>Methane Requirement (g CH4/g VC)</td>
<td>43</td>
</tr>
</tbody>
</table>

a. For the runs with VC toxicity, data are summarized for periods of operation for which this effect was minimized or eliminated.

b. Standard deviations in parentheses.

c. During period for which VC degradation kinetics were determined.

Efficiencies greater than 99.6% were observed in all four of these experimental runs (Table 3). The continuous-flow control run (without methanotrophs) exhibited less than 6% loss of VC concentration under these conditions (Fig. 2). Toxicity of VC was suspected to have limited the operation of the MAFEB system during parts of the continuous-flow runs with higher VC loading rates (runs 1 and 6). High concentrations of dissolved methane also appear to have inhibited VC degradation during some of the runs. These inhibitory effects are discussed later herein. After minimizing VC toxicity and methane inhibition, the fifth continuous-flow run resulted in the most successful operation of the system. During this reactor run, with an expanded-bed HRT of 5.5 h, the aqueous VC concentration was consistently reduced from about 2,000 μg/L to 2–3 μg/L throughout the run (Fig. 2). The VC removal efficiency during this run was 99.8%, and the average VC degradation rate was 10 mg VC/(L Eb d) or 1.3 mg VC/(g VS d) (Table 3), where the subscript eb equals expanded bed.

VC degradation rates and removal efficiencies were calculated for periods of steady operating conditions during each of the six continuous-flow runs in the MAFEB reactor (Table 3). VC degradation rates were observed to increase with increasing VC loading rate, and high degradation rates corresponded to high effluent VC concentrations (Fig. 3). VC degradation rates, expressed in terms of reactor expanded-bed volume, ranged from 7.6 to 20 mg VC/(L Eb d). On a biomass basis, the observed degradation rates varied from 0.96 to 2.5 mg VC/(g VS d). The operating effluent VC concentrations over which these rates were observed ranged from 3 to 140 μg/L (Table 3). All degradation rates were observed for low VC concentrations, under substrate-limited conditions, except for the runs for which VC toxicity effects were observed. Continuous-flow kinetics could not be determined for higher VC concentrations because of the apparent VC toxicity. This effect is discussed later.

The methane requirement for VC degradation during the continuous-
FIG. 3. VC Degradation Rates as Function of Effluent VC Concentration during Continuous-Flow Runs in MAFEB Reactor

flow tests varied from 40 to 240 g CH\textsubscript{4}/g VC (Table 3). Methane consumption was lowest during the two continuous-flow runs with the highest VC loading rates and effluent VC concentrations (1 and 6). The low methane consumption during these runs may have been related to suspected VC toxicity that would have lowered methane uptake rates. Methane consumption was the greatest during the runs with the lowest VC loading rates (Table 3). Since methanotrophic VC degradation is thought to be a cometabolic process, VC degradation might be expected to be related to methane consumption by a fixed cometabolism ratio. However, in the present experiments, no attempt was made to optimize (minimize) methane consumption, and the observed methane-use rates were probably much higher than any theoretical cometabolism ratio. Since VC degradation is possible in the absence of methane, it may be possible to optimize methane consumption to lower levels than those reported here.

**VC Toxicity/Inhibition**

Sustained aqueous VC concentrations between 5 and 10 mg/L were found to inhibit both the growth of the methanotrophic culture and its ability to oxidize VC. This inhibition was observed only in continuous-flow runs 1 and 6, during which high VC loading rates were employed and high effluent VC concentrations were sustained during part of the run. Influent and effluent VC concentrations during these two runs are shown in Fig. 4. VC toxicity or inhibition appeared to result in a poor VC removal efficiency during the beginning of each of these runs. In both cases, a reduction of the VC loading rate alleviated the problem, and VC removal efficiencies improved toward the end of the experiments (Fig. 4). Inhibition of the methanotrophs during these two runs was also characterized by a reduction in methane and oxygen use and an increase in effluent pH due to decreased
CO₂ production. Methane use rates decreased from about 2–4 L/(Lₑₑ d) at low VC operating concentrations to less than 0.6 L/(Lₑₑ d) with a VC operating concentration of 5 mg VC/L. VC inhibition reduced the observed daily growth rates by 80–90% and reduced observed yields by a factor of three to five (Table 2, compare runs 2–5 with runs 1 and 6). The inhibitory effect of VC was self-perpetuating because the reduction in VC degradation rates concomitantly resulted in a further increase in VC concentration and VC inhibition. The observed inhibition and/or toxicity may have been caused by toxic intermediates formed during VC degradation. Other researchers report that toxic epoxide intermediates formed during methanotrophic trichloroethene degradation were responsible for loss of methanotrophic activity (Alvarez-Cohen and McCarty 1991).

Recovery of the MAFEB system from VC inhibition was observed by turning off the feed to the reactor after a period of VC inhibition during run 6. The reactor was then operated in semibatch mode for a period of 10 days, during which time gas use gradually increased and the VC concentration gradually decreased. The VC concentration in the reactor was eventually reduced to 1 µg/L after 10 days (Fig. 4). This test demonstrated the ability of the culture to recover from the VC-induced stress, but the recovery was slow. This long recovery period indicates that the effect of VC may have been more than enzyme site saturation and may have involved some toxic effects.
Semibatch VC Degradation Kinetics

Short-term semibatch tests were conducted to further define VC degradation kinetics at higher VC concentrations and to investigate temperature effects and methane inhibition. Each semibatch test was conducted after a period of continuous operation during which steady growth, gas-uptake rates, and pH were maintained.

The effect of temperature on VC degradation rates was investigated by conducting semibatch runs in the MAFEB reactor at 35°C, 25°C, and 21°C (Table 4). Dissolved methane was maintained at low concentrations during these runs, usually less than 1 mg/L. During semibatch runs at all three temperatures, the measured aqueous VC concentration in the MAFEB reactor decreased exponentially from its initial value to near the detection limit (Fig. 5). At 35°C, the VC concentration decreased from an initial value of 5,400 μg/L to below detection limits (<5 μg/L) after 8 h. During the run at 25°C, the VC concentration decreased from 6,000 μg/L to 10 μg/L in 24 h. At 21°C the VC concentration decreased from an initial concentration of 6,000 μg/L to 130 μg/L in 14 h, and after 24 h, VC was undetectable in

TABLE 4. VC Degradation Rates and MAFEB Operating Parameter during Batch runs at 21°C, 25°C, and 35°C

<table>
<thead>
<tr>
<th>Operating Temperature</th>
<th>21°C (2)</th>
<th>25°C (3)</th>
<th>35°C (4)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reactor effluent pH</td>
<td>5.30</td>
<td>5.30</td>
<td>5.5</td>
</tr>
<tr>
<td>Expanded-bed volume (L,eb)</td>
<td>0.40</td>
<td>0.35</td>
<td>0.38</td>
</tr>
<tr>
<td>Total biomass content (g VS)</td>
<td>8.2</td>
<td>8.2</td>
<td>6.2</td>
</tr>
<tr>
<td>Initial VC concentration (mg/L)</td>
<td>5.7</td>
<td>3.4</td>
<td>4.3</td>
</tr>
<tr>
<td>Initial VC degradation rate (mg VC/g VD d)</td>
<td>25</td>
<td>31</td>
<td>63</td>
</tr>
<tr>
<td>(mg VC/L,eb-d)</td>
<td>530</td>
<td>680</td>
<td>970</td>
</tr>
</tbody>
</table>

FIG. 5. Semilog Plots of VC Concentration versus Time for Semibatch Runs at Three Operating Temperatures
the reactor effluent. The linear relationship between the log of the VC concentration and time indicates that VC degradation during these semibatch runs followed a first-order relationship over this concentration range.

Initial degradation rates during each run are listed in Table 4, both on a biomass basis and on the basis of expanded-bed volume. The highest VC degradation rate observed in these semibatch experiments was 63 mg VC/(g VS d), which was observed at 35°C. At 21°C, the maximum VC degradation rate was 25 mg VC/(g VS d). A linear relationship between degradation rate and concentration was observed at both temperatures over this concentration range (Fig. 6). The slope of the rate versus concentration plot for the 35°C data are about three times greater than that at 21°C, indicating a threefold increase in degradation rate with this 14°C degree increase in temperature. This effect of temperature roughly corresponds to a doubling of the degradation rate for every 10°C increase in temperature.

**METHANE INHIBITION**

Methane inhibition was suspected to be limiting VC degradation during the continuous-flow runs because high dissolved methane concentrations correlated with decreased VC degradation rates. In all of the continuous-flow runs, the most efficient VC degradation was observed with low dissolved methane concentrations, while the least efficient VC degradation was observed with high dissolved methane concentrations. Degradation of VC to concentrations below 10 µg/L was only observed when the dissolved methane concentration above the bed was below about 0.5 mg/L. It appears that methane inhibits the ability of the MMO enzyme to oxidize VC. Such inhibition has been recently reported for methanotrophic TCE degradation (Henry and Grbic-Galic 1991).

Methane inhibition was examined further by conducting semibatch runs in the MAFEB reactor with a series of controlled dissolved methane concentrations. Stable dissolved methane concentrations were maintained in
four semibatch runs by adjusting gas recycle rates, resulting in dissolved methane concentrations above the expanded bed ranging from 0.2 to 8.5 mg/L. The observed VC degradation rates were approximately first order with VC concentration, and were strongly influenced by dissolved methane in the reactor (Fig. 7). Rates up to 15 mg VC (g VS d) were observed with low dissolved methane concentrations, while little or no VC degradation was observed when the dissolved methane concentration was increased to 8.5 mg/L. Methane inhibition of VC degradation can be summarized by comparing VC degradation rates measured at a time during each semibatch run when the operating VC concentration was 1 mg/L. These rates are
plotted as a function of dissolved methane concentration in Fig. 8. VC degradation decreased linearly with increasing dissolved methane concentration up to a concentration of about 2 mg CH$_4$/L. Above 2 mg CH$_4$/L, VC degradation was extremely inhibited (Fig. 8).

CONCLUSIONS

These experiments have demonstrated the feasibility methanotrophic VC degradation in attached-film expanded-bed bioreactors on a laboratory scale. Under optimized conditions (low dissolved methane concentration and low VC concentration), VC removal efficiencies as high as 99.8% were observed in the MAFEB system on a continuous-flow basis. Vinyl chloride concentrations were reduced from 2,000 µg/L to 3 µg/L with a hydraulic retention time in the bed of 5.5 h. The methane requirement for VC degradation ranged from 40 to 240 g CH$_4$/g VC, with the higher values associated with low VC loading rates. Vinyl chloride degradation rates during continuous-flow operation varied from 10 to 20 mg VC/(L$_{eb}$ d) for operating VC concentrations between 1 and 100 µg VC/L.

Much higher VC degradation rates were observed during batch runs than during continuous-flow runs because the batch runs were conducted with much higher VC concentrations than the continuous-flow runs. The VC concentration employed in continuous-flow experiments was limited because high VC loading rates apparently resulted in toxic effects. The continuous-flow runs were substrate limited with respect to VC, and they were conducted with extremely low effluent VC concentrations, often as low as 3 µg/L. Under these conditions, VC degradation rates of about 1 mg VC/(g VS d) were observed. During semibatch runs, VC degradation rates were observed to follow a first-order relationship with VC concentration over a range of 0 to 5 mg VC/L. This first-order relationship would predict much lower continuous-flow VC degradation rates at the operating VC concentrations employed in the continuous-flow runs than were actually observed. Further studies are needed to clarify the large differences between kinetics under semibatch and continuous-flow operations. Continuous-flow kinetics represent a better approximation of VC degradation for a practical treatment system that would presumably operate continuously. However, these data suggest that some process modification might achieve more rapid removals, such as sequencing semibatch operation.

The VC degradation rates observed during semibatch runs were similar to those reported previously by Fogel et al. (1987) for VC degradation by a suspended methanotrophic consortium. These researchers reported a degradation rate of 14.6 nmol/(mg h) [22 mg/(g d)] at a concentration of 1 to 2 mg VC/L. This reported rate is similar to rates observed during semibatch runs in the present study. It is encouraging that VC degradation rates reported for laboratory microcosms could be maintained in a bioreactor.

Two important limitations of methanotrophic VC degradation were identified during the course of these experiments. The first is the inhibitory or toxic effect of VC on the methanotrophs, and the second is methane inhibition of methanotrophic VC degradation. Continuous exposure of the attached biomass to VC concentrations of 5 to 10 mg/L, at VC loading rates of 60 mg VC/(L$_{eb}$ d), severely limited the metabolism of both methane and VC. However, since the MAFEB reactor operates as a completely mixed system, VC toxicity should not cause system failure as long as conditions are maintained in the reactor that continually support VC degradation so that effluent VC concentrations are kept below 5 mg/L. However, it is also
possible that toxic intermediates formed during methanotrophic VC degradation result in cell death proportional to the amount of VC degraded. Other researchers have suggested that this is the case for TCE degradation, and the amount of TCE degraded per amount of cells killed was termed the “transformation capacity” (Alvarez-Cohen and McCarty 1991). Other researchers reported on the same phenomenon and defined an “inactivation constant,” which is the amount of cells killed per amount of VC degraded (Oldenhuis et al. 1991). If this is the case for VC degradation, then loading rates must be maintained in accordance with growth rate to avoid system failure.

The second factor, methane inhibition, was found to limit VC degradation when the dissolved methane concentration in the reactor was above 0.5 mg/L. Methane inhibition was avoided in continuous-flow runs by reducing the methane gas recycle rate to maintain low partial pressures of methane in the diffuser headspaces. This adjustment may prove more difficult with a full-scale system, and in that case alternative strategies would be required for maintaining low dissolved methane concentrations. An example of such a strategy would be to employ a sequencing semibatch treatment system in which VC and methane feeds are alternately pulsed to the reactor system. Preliminary experiments have indicated that the expanded-bed system continues to degrade VC for at least 6 h after eliminating the methane supply. Operation in this mode would allow VC degradation to be executed in the absence of inhibiting methane. After several hours in this mode, the MAFEB system would be rejuvenated with methane off-line.

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