**250** Scale up and mass balance of affinity purification of native  $\beta$ -lactoglobulin. Harit K. Vyas<sup>\*</sup>, J. M. Izco, and R. Jimenez-Flores, *Dairy Products Technology Center, Cal Poly.* 

The process of affinity purification of  $\beta$ -lactoglobulin in its native form using all-trans-retinal immobilized on Celite R-648<sup>TM</sup> was scaled up and applied to fractionate industrial sweet whey. Three different ways of mixing the Celite  $R-648^{TM}$  and whey for the interaction between alltrans-retinal and  $\beta$ -lactoglobulin were tried at pilot scale. The three methods used were (1) a continuous operation using a column packed with Celite  $R-648^{TM}$ , (2) a batch operation in a stirred tank and (3) a continuous operation using a fluidized Celite R-648<sup>TM</sup> column. Adsorption and desorption of  $\beta$ -lactoglobulin were carried out at pH 5.1 and 7.0, using 0.01 and 0.1M phosphate buffers, respectively. The phosphate buffer containing desorped  $\beta$ -lactoglobulin was concentrated 20 times using ultrafiltration and then freeze-dried. The packed column, stirred tank and fluidized column produced  $\beta$ -lactoglobulin with purity of 80, >95 and >95%, and recovery of 0.65, 2.88 and 2.88g per kg of Celite R-648<sup>TM</sup>, respectively. The comparative poor purity and recovery of  $\beta$ -lactoglobulin in the case of the packed column was attributed to insufficient contact between the passing fluids and the Celite  $\mathrm{R\text{-}648^{TM}}$ during adsorption, desorption and intermittent washing. The fluidized column method being a continuous operation with a gentle mixing action, was considered the best suited for further scale up to the industrial level.

Key Words: Process scale up, affinity purification,  $\beta$ -lactoglobulin